







Review

The Potential of Nutraceutical Supplementation in Counteracting Cancer Development and Progression: A Pathophysiological Perspective

Carmen Altomare, Roberta Macrì, Maria Serra, Sara Ussia, Giovanna Ritorto, Jessica Maiuolo, Carolina Muscoli, Enzo Perri and Vincenzo Mollace

Special Issue

Effects of Plant Extracts on Human Health—2nd Edition

Edited by

Prof. Dr. Fátima Regina Mena Barreto Silva and Prof. Dr. Diana Marcela Aragon Novoa









Review

The Potential of Nutraceutical Supplementation in Counteracting Cancer Development and Progression: A Pathophysiological Perspective

Carmen Altomare ¹, Roberta Macrì ¹,* ¹, Maria Serra ¹,* ¹, Sara Ussia ¹, Giovanna Ritorto ¹, Jessica Maiuolo ², Carolina Muscoli ¹, Enzo Perri ³ and Vincenzo Mollace ¹

- Pharmacology Laboratory, Institute of Research for Food Safety and Health IRC-FSH, Department of Health Sciences, University Magna Graecia of Catanzaro, 88100 Catanzaro, Italy; carmen.altomare@studenti.unicz.it (C.A.); saraussia1598@gmail.com (S.U.); giovanna.ritorto@studenti.unicz.it (G.R.); muscoli@unicz.it (C.M.); mollace@libero.it (V.M.)
- ² Laboratory of Pharmaceutical Biology, IRC-FSH Department of Health Sciences, University "Magna Græcia" of Catanzaro, 88100 Catanzaro, Italy; maiuolo@unicz.it
- Research Centre for Olive, Fruit and Citrus Crops, Council for Agricultural Research and Economics, 87036 Rende, Italy; enzo.perri@crea.gov.it
- * Correspondence: roberta.macri@unicz.it (R.M.); maria.serra@studenti.unicz.it (M.S.)

Abstract

Cancer is a major cause of morbidity and mortality across the globe, with a substantial increase in cases anticipated over the next few decades. Given the constraints and adverse effects associated with standard cancer therapies, the contribution of diet and nutraceuticals to cancer prevention and treatment is receiving increased scrutiny. A diet rich in plant-based foods, extra virgin olive oil (EVOO), and bioactive compounds, including the Mediterranean Diet, has been associated with reduced cancer risk and improved treatment outcomes. This review aims to explore the complex mechanisms of the MedDiet and nutraceuticals (polyphenols, flavonoids, terpenoids) in cancer prevention, to determine their potential as cancer treatment adjuvants. Promising results show that key compounds such as bergamot polyphenolic fraction (BPF), cynaropicrin, oleuropein, quercetin, resveratrol, and serotonin can modulate oxidative stress, inflammation, the tumor microenvironment, the cell cycle, and drug resistance. A significant observation is that many of these substances demonstrate dual dose-dependent activity; they function as antioxidants in healthy cells but induce pro-oxidant and pro-apoptotic effects in cancerous cells. Their ability to boost chemotherapy's effectiveness and safety while lessening side effects and offering combined advantages is also explored. To summarize, this review suggests that the Mediterranean Diet and nutraceutical supplements may help prevent and manage cancer, but more research is needed to confirm their benefits.

Keywords: Mediterranean Diet; nutraceuticals; dietary polyphenols; antioxidant compounds; oxidative stress; inflammation; cancer prevention; health-promoting phytochemicals



Academic Editors: Diana Marcela Aragon Novoa and Fátima Regina Mena Barreto Silva

Received: 18 June 2025 Revised: 14 July 2025 Accepted: 15 July 2025 Published: 18 July 2025

Citation: Altomare, C.; Macrì, R.; Serra, M.; Ussia, S.; Ritorto, G.; Maiuolo, J.; Muscoli, C.; Perri, E.; Mollace, V. The Potential of Nutraceutical Supplementation in Counteracting Cancer Development and Progression: A Pathophysiological Perspective. Nutrients 2025, 17, 2354. https://doi.org/10.3390/nu17142354

Copyright: © 2025 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Cancer is the second leading cause of death globally, trailing only heart disease. According to the International Agency for Research on Cancer (IARC), there were about 20 million new cancer diagnoses and 9.7 million deaths globally. By 2050, annual cases could hit 35 million, assuming current incidence levels persist [1]. Several factors, including lifestyle, daily routines, and diet, contribute to the rising cancer rates [2]. Epidemiological

Nutrients **2025**, 17, 2354 2 of 41

and clinical research shows a link between nutrition and the development or progression of various cancers (colon, breast, and prostate), classifying them as diet related [3]. The beneficial effects of a healthy lifestyle, and its correlation with the Mediterranean Diet's (MedDiet) wide range of health benefits, are supported by several studies [4], including reduced risk of cancer and cardiovascular disease (CVD), reduced metabolic risk of diabetes mellitus (DM), obesity, and metabolic syndrome (MS), increased longevity, and improved cognitive function. Furthermore, the MedDiet is linked to a healthy gut microbiome, impacting cancer development and treatment.

The hallmark of cancer is the conversion of some normal cells into malignant neoplastic cells [5].

Therefore, uncontrolled proliferation, evasion of growth suppressors and the immune system, angiogenesis, metastasis, and metabolic reprogramming are the main characteristics [6]. Cellular and subcellular microenvironments and genetic instabilities are key to this multistep process. Malfunction of the core cell cycle leads to unstable deoxyribonucleic acid (DNA) and mutations, furthering tumorigenesis [7]. The high number of cancer cases urgently requires new treatment approaches to address the weaknesses of traditional methods like surgery, hormone replacement therapy (HRT), chemotherapy, and radiation, which often cause drug resistance and harmful side effects [8]. Because they boost tumor response to treatments and decrease systemic toxicity, the wide array of natural compounds in the MedDiet are a potential cancer-fighting strategy [9]. This review aims to highlight the effects of natural compounds, as nutraceuticals, in the management of cancer therapy. Due to their proven antioxidant and inflammatory properties, as well as their ability to modulate the cell cycle, natural compounds—in particular extracts from a selection of foods characteristic of the MedDiet, such as Citrus bergamia Risso & Poiteau, Cynara cardunculus L., Olea Europaea L., foods rich in quercetin, resveratrol (RV), as well as Mediterranean plants such as Ferula communis L.—may be relevant for synergism with chemotherapeutic drugs while reducing their related toxic effects.

1.1. Complexities of Cancer Pathogenesis

Cancer development is a complex process influenced by various factors, leading to diverse clinical presentations, and various factors contribute to its pathogenesis, each a potential therapeutic target (e.g., chemicals, viruses, genetic, and non-genetic factors).

Several studies have shown that carcinogenesis is related to somatic mutations of stem cells, which generate a subpopulation of cancer stem cells (CSCs) with self-renewal capabilities, producing daughter cells with highly tumorigenic and invasive potential, originating primary tumors [10]. Evidence suggests that epigenetic mutations can provide tumor cells with a reproductive advantage over normal cells, leading to impaired tumor suppression and uncontrolled tumor growth [11]. However, the inherent adaptability of healthy cells, allowing epigenetic and phenotypic shifts leading to CSC formation, is pivotal in cancer development [12]. Oncogene and tumor suppressor alterations in this scenario accelerate cell growth and may cause a more aggressive metastatic phenotype.

The occurrence of Rat sarcoma virus (RAS) and its family genes, Kirsten rat sarcoma viral oncogene homolog (KRAS) (*KRAS4A* and *KRAS4B*), Neuroblastoma RAS viral oncogene homolog (NRAS), or Harvey Rat sarcoma virus (HRAS) [13] are among the most dysregulated protooncogenes and best characterized. Oncogenic activation of RAS results in ligand-independent hyperstimulation of downstream signaling cascades, including the extracellular signal-regulated kinase 1/2 (ERK1/2) [14] and phosphatidylinositol 3-kinase (PI3K) promoting cell growth, cell cycle entry, cell survival, and cell adhesion. Moreover, mutations in KRAS lead to chemoresistance mechanisms [15]. The function of tumor protein p53 (p53), a tumor suppressor, is widely understood. p53 gene mutations disrupt the

Nutrients 2025, 17, 2354 3 of 41

normal response to DNA damage, blocking apoptosis and cell cycle arrest, thus causing abnormal cell division. The development of cancer is linked to the epithelial-mesenchymal transition (EMT) and a metabolic shift from glycolysis to oxidative phosphorylation [16]. Epithelial cells undergoing EMT lose apical-basal polarity and cell-cell junctions, gaining mesenchymal characteristics, such as invasion and migration. In addition, cells develop stem cell characteristics and become resistant to chemotherapy [17]. Signaling pathways to activate and maintain EMT include notch signaling pathway (NOTCH), wingless-related integration site (Wnt), glycogen synthase kinase 3 beta (GSK-3β), and transforming growth factor beta $(TGF-\beta)$ [18]. These pathways lead to loss of cell-cell adhesion and increased migratory cell ability by activating transcription factors including Slug, Twist, Snail, or Zinc finger E-box binding homeobox 1 and Zinc finger E-box binding homeobox 2 (ZEB1/2) [19], which in turn repress epithelial markers such as E-cadherin. Cancer cells metabolism shift from production energy by glycolysis to oxidative phosphorylation, mediated by pathways such as PI3K/Akt/mTOR and AMP-activated protein kinase (AMPK) [20,21]. This network encompasses cancerous and non-cancerous host cells, alongside the modified extracellular matrix (ECM) and vascularization. Tumor cell invasion is fueled by the tumor microenvironment (TME), a key factor in tumor development and resistance to treatment. Its inflammatory, hypoxic nature drives adaptive pathways, enzyme overexpression, high adenosine triphosphate (ATP), and low pH, resulting in acidity [22]. The TME includes immune cells from both innate and adaptative systems, endothelial cells (ECs), cancer-associated fibroblasts (CAFs), pericytes, adipocytes, and other cell types that vary by tissue. In the TME, there are also myeloid-derived suppressor cells (MD-SCs), tumor-associated macrophages (TAMs), and regulatory T-cells (Tregs), which secrete immunosuppressive molecules such as transforming growth factor beta (TGF-β) or modulating immune checkpoint [23]. Furthermore, the tumor tissue's commensal microbiome is a key TME component, affecting tumor development and growth. The distinct intratumoral and intracellular microbial composition of different tumor types allows for the correlation of microbial metabolic pathways and drug treatment characteristics. Moreover, dysbiosis can trigger a persistent inflammatory immune response, via the microbial activation of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κβ), a cancer-linked inflammation regulator, thus promoting tumor growth [24].

1.2. Nutrition and Nutraceutical Compounds in Cancer Risk Modulation

Diet, nutrition, and lifestyle are correlated with cancer incidence, progression, and treatment response in some epidemiological studies. Age-related cardiovascular and metabolic diseases are also linked, according to scientific evidence. Human and animal studies demonstrate a link between diet, epigenetic changes, and cancer risk. It is possible to assert that lifestyle and diet, and particularly the MedDiet model, play a key role as a modulator of cancer risk. It represents not only a diet but also a lifestyle model in the Mediterranean area, where the incidence of several types of cancer is lower than in other areas [25]. Such incidence is associated with a potential protective role of the MedDiet on human health [26]. The plan emphasizes plenty of vegetables, fruits, legumes, whole grains, and nuts. The diet features extra virgin olive oil (EVOO) as its main fat, plus regular fish, moderate animal protein, and red wine.

The MedDiet has a high intake of B vitamins (vitamins B1, B2, niacin, B6, folate or B12), antioxidant vitamins (vitamins E and C), minerals (iron, selenium, phosphorus, and potassium), and fiber. A variety of nutraceuticals included in the MedDiet have shown an inverse association with CVD, metabolic disease, neurodegenerative disease, and various types of cancer, both directly and through epigenetic mechanisms [27]. Strict adherence to the MedDiet is associated with a lower risk of multiple diseases thanks to phytochemicals

Nutrients **2025**, 17, 2354 4 of 41

("non-nutrient" components) present in its constituent foods, such as polyphenols, phytosterols, carotenoids, monounsaturated fatty acids (MUFAs) and polyunsaturated fatty acids (PUFAs) [28]. Specifically, polyphenols, water-soluble phenylpropanoids, contain at least one aromatic ring and a benzene ring with hydroxyl substituents. Plants synthesize these secondary metabolites to cope with stress like pathogen attacks, ultraviolet radiation (UV) radiation, or harsh weather. They are classified as: (i) flavonoids, characterized by the formula C6-C3-C6, among which there are flavonols, flavanols, flavones, flavanones, isoflavones, and anthocyanins; (ii) non-flavonoids, characterized by only one phenolic ring, including phenolic acids (hydroxybenzoic acids such as vanillic acid and gallic acid) and cinnamic acids (ferulic and caffeic acids), lignans, tannins, and stilbenes, including RV [29]. Foods contain various polyphenols such as flavan-3-ols, isoflavones, flavonols, flavanones, flavones, anthocyanins, and stilbenes, impacting color, taste, smell, bitterness, astringency, and resistance to oxidation. The range of polyphenol levels is widely conditioned by environmental conditions and soil type [30]. Studies demonstrate the relationship between the structural chemical conformation of polyphenols and their biological actions, affecting bioavailability and absorption [31]. In fact, the complex nature of polyphenols results in poor small intestinal absorption. Following absorption, liver enzymes (glucuronidation, sulfonation, methylation) modify phenolic substances before systemic circulation and tissue distribution. The inability of complex phenols to be absorbed leads to their biotransformation by the gut microbiota into smaller, bioavailable derivatives [32]. Epidemiological studies suggest that sufficient polyphenol intake, from food or supplements, may reduce oxidative stress, chronic inflammation, and cancer risk [33]. Moreover, increased polyphenol intake enhances cardiometabolic health via anti-inflammatory, antioxidant, vasodilatory, high-density lipoprotein (HDL)-raising actions, and by inhibiting low density lipoprotein (LDL) oxidation [34]. Polyphenols have been shown to reduce blood glucose levels by mechanisms including the inhibition of glucose uptake in the gut and other tissues [35]. Indeed, epidemiological studies also demonstrate that polyphenols protect against Metabolic Syndrome, a combination of hypertension, dyslipidemia, insulin resistance, and central obesity that contributes to cardiovascular and metabolic diseases [36]. Polyphenols can restore redox balance [37], impacting arachidonic acid metabolism through the inhibition of various enzymes, such as phospholipase A2 (PLA2), lipoxygenase (LOX), and cyclooxygenases (COX-1 and COX-2); consequently, there is a reduction in the release of prostaglandins, thromboxanes, leukotrienes, and other mediators of inflammation [38]. Metal ion chelation mechanisms enable these molecules to exert their chemoprotective effects, including cell cycle arrest induction, pro-apoptotic process induction, and immune system regulation [39]. Polyphenols have both estrogenic and antiestrogenic effects, impacting how anticancer drugs are processed in the body, thus reducing their toxicity [40]. In addition, some polyphenols have been shown to be able to disrupt the elongation of telomeric ends, which could have an important impact on cancer cell replication, and they are effective as anti-aging compounds [41]. In fact, the level of antioxidants in plasma depends on the consumption of antioxidant-rich foods and supplements, such as those containing polyphenols. Natural compounds, by modulating specific proteins, can regulate drug resistance, which occurs through several mechanisms, including enhanced DNA damage repair, invasiveness, altered drug efflux and metabolism, modification of drug targets and TME [42]. The important role of nutraceuticals in oncology has led to them being called onconutraceuticals [43]. In cancer treatment, their application focuses on discovering synergistic effects with anticancer drugs to improve treatment success, decrease side effects (e.g., cardiotoxicity, hepatotoxicity), and overcome drug resistance. Table 1 generated from the literature data, presents the principal biological properties of the key nutraceuticals of

Nutrients **2025**, 17, 2354 5 of 41

the MedDiet and Figure 1 shows the molecular diagrams of the main natural compounds discussed in this review.

 Table 1. The Mediterranean Diet, nutraceuticals, and their biological properties.

Nutraceuticals	Bioactive Compounds	Main Sources	Biological Effects	References
Citrus bergamia Risso & Poiteau extract	Naringin, neohesperidin, neoeriocitrin, C-glucoside, flavanone O-glycosides, rhoifolin, 40-O-glucoside, neodiosmin, rhoifolin, poncirin limonene, linalool, linalyl acetate	Bergamot juice (BJ), bergamot essential oil (BEO)	Hypolipemic, hypoglycaemic anti-inflammatory, antioxidant, anticancer	[44–47]
Cynara cardunculus L. extract	Cynaropicrin (cyn), chlorogenic acid, dicaffeoylquinic acids, luteolin, inulin	Leaves, flowers, roots, by-products	Hepatoprotective, antioxidant, antimicrobial, antiobesity, chemopreventive [48,49]	
Olea Europaea L. extract	Hydroxytyrosol, oleuropein, tyrosol, oleic acid, omega-3, omega-6	Olive oil (OO), olive leaves, Olive mill wastewater (OMWW)	Antioxidant, cardioprotective, antitumoral, anti-inflammatory, anti-aging, antibacterial, prevention of metabolic disorders and chronic diseases	[50–53]
Quercetin	Flavonol (free and glycosylated forms)	Onions, apples, berries, broccoli, tea, cherries, tomatoes, asparagus, peas, grapes, coriander seeds	Antioxidant, anti-inflammatory, antiproliferative, antiviral, cardioprotective, antiaging, prevention of metabolic disorders, chronic diseases, platelet aggregation, lipid peroxidation, and capillary permeability, modulating the composition of the gut microbiota	[54,55]
Resveratrol	Trans-RV	Grapes skin, red wines, blueberry, cranberry, peanuts, bilberry	Antioxidant, anti-inflammatory, pro-apoptotic, antitumor, telomerase inhibition, improves gut barrier	[30,56–58]
Ferula Communis L. extract	Ferutinin, sesquiterpenes, ferulenol, ferulone A and B, flavonoids	Roots, rhizomes, latex	Selective estrogen receptor modulator (SERM-like), antiproliferative, dose-dependent estrogenic effect; antioxidant effects, antidiabetic, antimicrobial, cytotoxic actions	[59,60]

BJ: bergamot juice; BEO: bergamot essential oil; Cyn: cynaropicrin; OO: olive oil; OMWW: olive mill wastewater; RV: resveratrol; SERM-like: selective estrogen receptor modulator.

1.2.1. Bioactive Compounds from Citrus bergamia Risso & Poiteau and Cynara cardunculus L.

Citrus bergamia Risso & Poiteau, commonly referred to as "Bergamot", is a Mediterranean citrus fruit, and is considered an endemic plant of the Calabria region. In fact, while its precise botanical and geographical origins are unknown, bergamot is cultivated along the

Nutrients **2025**, 17, 2354 6 of 41

Calabrian Ionian coast, between Bianco and Reggio Calabria [44]. This plant belongs to the Rutaceae family, Esperidea subfamily. Three varieties of bergamot are known—"Fantastico", "Femminello", and "Castagnaro". The relevance of this plant stems from the biological role of herbal preparations, particularly BEO and BJ. BEO is a volatile oil obtained by scraping and cold pressing the fruit peel; it contains up to 93-96% volatile compounds (limonene, linalool, and linalyl acetate), while the remaining percentage is a variable fraction of nonvolatile substances (pigments, waxes, coumarins, and psoralens) [45]. BJ is derived from the endocarp and pulp of bergamot fruits, featuring a distinctive composition of compounds also present in albedo, including flavonoids like naringin, neohesperidin, and neoeriocitrin, as well as C-glucoside, flavanone O-glycosides, and flavone O-glycosides such as rhoifolin 40-O-glucoside, neodiosmin, rhoifolin, and poncirin, along with furanocoumarins like bergapten and bergamottin. The major biologically active polyphenols from BJ were found in the bergamot polyphenolic fraction (BPF). High-dose toxic furanocoumarins are removed from the BJ through an alkalinization step in production [61]. The key attraction of bergamot is the diverse health benefits stemming from the pharmacological activity of its components. Furthermore, the recent circular economy approach has highlighted the value of bergamot waste, supporting its use in therapies and advancing scientific understanding [46,47].

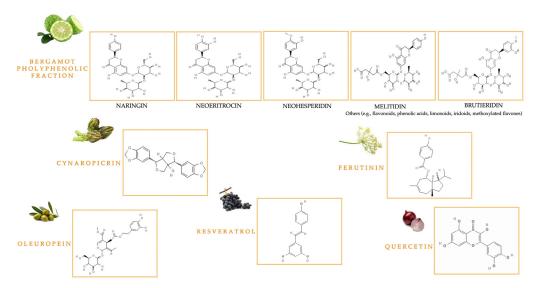


Figure 1. Diagrams of molecular structures.

Cynara cardunculus L., commonly referred to as "cardoon," represents a perennial Mediterranean plant belonging to the Cardueae tribe Cass. (Cynarae Less.), Asteraceae family, which includes three botanical taxa, the artichoke (C. cardunculus L. var. scolymus L. Flowers), the perennial wild thistle (C. cardunculus L. var. sylvestris (Lamk) Flowers), and the domestic thistle (C. cardunculus L. var. altilis DC.). This Mediterranean plant thrives despite harsh conditions, including extreme temperature swings and poor soil [62]. The immature flower structure of the artichoke, encompassing the leaves (bracts) and upper receptacle, is what is edible. Due to its nutritional profile—high in minerals, vitamins, amino acids, proteins, and low in lipids—cardoon qualifies as a functional food. The roots and rhizomes are rich in inulin, a crucial carbohydrate contributing to its high nutritional value and supporting gut microbiota [49], while cardoon leaves are a source of antioxidants such as dicaffeoylquinic acids and luteolin. Inedible plant matter accounts for 80–85% of plant biomass, resulting in wasted bioactive substances. Thus, the byproduct use of non-food parts yields useful pharmacologically active molecules such as polyphenols and flavonoids. Chlorogenic acid (5-O-caffeoylquinic acid) and cynaropicrin, a sesquiterpene

Nutrients 2025, 17, 2354 7 of 41

lactone, are responsible for cardoon's bioactivity, including hepatoprotective, antioxidant, chemopreventive, antiobesity, and antimicrobial properties [48].

1.2.2. Bioactive Polyphenolic Compounds in Olive Oil

Olive Oil (OO) represents the most important product obtained from the evergreen tree in the Mediterranean basin Olea Europaea L., a key component of the MedDiet. The process uses only water and room temperature to crush the olives during malaxation, creating the product without chemical solvents. The biological activity of Olive Oil is largely due to its rich phytochemical composition, most notably the lipophilic fraction (95– 97%) of MUFAs (e.g., oleic acid) and PUFAs (e.g., omega-3 and omega-6). The other fraction consists of tocopherols, phytosterols, squalene, and phenols including oleuropein, tyrosol, and hydroxytyrosol [63,64]. The European Food Safety Authority (EFSA) has determined that "phenols in Extra Virgin Olive Oil (EVOO) contribute to the protection of blood lipids from oxidative stress," and this claim can be added to a product label when it contains at least 5 mg of hydroxytyrosol and its derivatives (e.g., oleuropein) per 20 g. Preclinical data indicates that oleuropein and hydroxytyrosol offer cardioprotection via mechanisms, including oxidative stress reduction, anti-inflammation, and homocysteine/cholesterol level modulation [50]. EVOO's unique phenolic profile, shaped by factors like cultivar and processing, is vital to its role in preventing a range of diseases, from Metabolic Syndrome to cancer [51,52]. Furthermore, Olive Mill Wastewater (OMWW) a byproduct from industrial EVOO processing, contains a high concentration of bioactive polyphenolic compounds (e.g., oleuropein) [53].

1.2.3. The Key Bioactive Role of Quercetin and Resveratrol

Quercetin and resveratrol, bioavailable polyphenols with antioxidant, anti-inflammatory, antiviral, and antiproliferative properties, are commonly found in foods characteristic of the MedDiet. Specifically, the flavonol quercetin (2-(3,4-dihydroxyphenyl)-3,5,7-trihydroxychromen-4-one) is prevalent in nature; key dietary sources include fruits and vegetables such as apples, berries, and onions [65]. Plants contain quercetin as glycosides (with attached sugars) or aglycones (without attached sugars), but the most common type, quercetin-3-O-glucoside, is hydrophilic and thus poorly absorbed. Consequently, quercetin demonstrates very low oral bioavailability. Extensive research focuses on this plant compound due to its effects on metabolic disorders, eye diseases, arthritis, and CVD [54,55].

Resveratrol is a stilbene and is produced by UV irradiation or fungal infection. However, antitumor, antioxidant, and anti-inflammatory activities are due to the trans-form of this bioactive compound [56], whereas there is currently only limited evidence for the cisisomer [66]. RV is found in various nutritious foods, including grape skins and seeds, and is frequently detected in red wine, blueberries, cranberries, peanuts, and bilberries. The glycosylation of RV in food enhances its stability and bioavailability by preventing enzymatic oxidation [67]. The nutraceutical potential of quercetin and RV is being actively researched due to their effects on inflammation, gut microbiota, oxidative stress, and cancer [57,58].

1.2.4. Bioactive Compounds from Ferula communis L.

The genus *Ferula communis* L. subsp. *communis* (Giant fennel) belongs to the *Apiaceae* family and is an herbaceous perennial plant containing latex. Central and South-West Asia, North Africa, and the Mediterranean region are home to approximately 180 species. Two distinct chemotypes have been identified, exhibiting different biological effects: a poisonous type with prenylated coumarins, and a non-poisonous type with sesquiterpene daucane esters. In addition, in plants of the genus *Ferula*, oleo-gum-resins are also found [68]. They are defined by many natural bioactive compounds, primarily from roots, leaves, and

Nutrients 2025, 17, 2354 8 of 41

rhizomes, including sesquiterpenes, sesquiterpene lactones, flavonoids, coumarins (like ferulenol), coumarin esters (like ferulone A, B), and phytoestrogens (like ferutinin) [69]. The non-toxic chemotype's phytoestrogens make it estrogenic—high extract concentrations can thus create a doubled ferutinin dose–response; unlike the other chemotype, which is toxic and causes severe hemorrhagic intoxication from the ingestion of its aerial parts leading to potentially lethal "ferulosis" [70]. The toxicity is due to the prenylated coumarin ferulenol; removing it makes the extracts non-toxic since the remaining prenylated coumarins are not toxic at their concentrations. Several *in vivo* and *in vitro* studies reported its pharmacological properties, including antidiabetic, antimicrobial, antiproliferative, and cytotoxic actions [59, 60]. In vitro studies and preclinical models have shown that ferutinin can be considered a selective estrogen receptor modulator (SERM), but its mechanism of action is not totally clear compared with classical SERMs. Its action on estrogen receptors depends on cell type and concentration; it is an estrogen receptor alpha (ER α) agonist and may act as an estrogen receptor beta $(ER\beta)$ agonist or antagonist. Therefore, the effects of ferutinin on gene expression and cell signaling pathways within estrogen-sensitive cancers (e.g., ERpositive breast cancer (BC) and endometrial cancer (EC)) are dose-dependent [71]. Further studies of this typical Mediterranean plant need to explore the underlying mechanisms and potential pharmaceutical applications.

2. Biological Functions and Pathological Implications of Reactive Oxygen and Nitrogen Species

Reactive oxygen/nitrogen species (ROS and RNS) are unstable, thus highly reactive and partially reduced, oxygen- and nitrogen-derived molecules that include superoxide anion (O_2^-) , hypochlorous acid (HOCl), hydroxyl radical, peroxyl radicals, nitric oxide, and peroxynitrite. The presence of a free electron on the oxygen atom allows oxygenated free radicals to react with sugars, proteins, and lipids; among free radicals, the superoxide anion is the most prevalent: oxidative phosphorylation in mitochondria, along with activated phagocytes, produces it via electron leakage from the electron transport chain to molecular oxygen [72].

Primarily, superoxide (O_2^-) will dismutate into oxygen and hydrogen peroxide (H_2O_2) . Although superoxide is unreactive with DNA, H₂O₂, in the presence of iron or copper, can become highly reactive hydroxyl radicals (•OH) through the Fenton reaction. The generation of ROS, including this process and superoxide-derived ROS, is linked to various diseases, including cancer, cardiovascular diseases, and neurodegenerative disorders [73,74]. In hepatocytes and macrophages, ROS are produced during processes like aerobic respiration and inflammation. ROS signaling, regulated by molecules and post-translational modifications (PTMs), triggers specific cellular responses based on ROS generation's nature and duration [75]. Cell differentiation and apoptosis, induced by ROS, contribute to physiological aging. RNS include nitric oxide and peroxynitrite: these molecules readily oxidize, thus damaging DNA and other biomolecules. Oxidative stress biomarkers are ROS-production enzymes such as nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, myeloperoxidase, xanthine oxidase (XO), and endothelial nitric oxide synthase (eNOS). Well known biomarkers of ROS-induced chemical modifications are the advanced glycation end products (AGEs), the oxidized low-density lipoprotein (OxLDL), and malondialdehyde (MDA) [76]. Therefore, a crucial factor is keeping the body's oxidative state balanced by generating and eliminating free radicals properly [77].

Indeed, high cellular ROS levels damage intracellular components like DNA, causing mutations and cell transformation [78]. Current findings suggest a link between ROS production and both internal and external influences. The electron transport chain within mitochondria is the primary endogenous source of ROS. High ROS levels cause DNA

Nutrients **2025**, 17, 2354 9 of 41

damage, oncogene activation, and tumor growth, whereas moderate levels induce cell death or senescence, thus suppressing tumor development [79].

Exogenous sources of ROS involve an unbalanced diet, anticancer therapy, radiation, smoking, drugs, and alcohol [80]. Chemotherapy drugs, including anthracyclines (doxorubicin (Dox), daunorubicin (Daun)), and camptothecins can cause various complications in the CV system due to ROS [81]. This is related to an increased lipid peroxidation and reduced levels of antioxidant molecules such as glutathione (GSH) [82]. It represents a non-enzymatic antioxidant, along with vitamins A, C, and E, and flavonoids. Enzymatic antioxidants include glutathione peroxidase (GPX), glutathione reductase (GRX), catalase (CAT), superoxide dismutase (SOD), superoxide reductase, thyroxine (TRX), and peroxiredoxin (PRX). The GSH system involves GSH, GR, glutathione transferase (GST), and GPX. Its deficiency or an altered GSH/GSSH ratio can lead to inflammation and tumor progression due to the occurrence of oxidative stress [83]. Dietary antioxidants are crucial, as evidenced by research on the diverse array of antioxidant systems [84,85]. By modulating oxidative stress, natural compounds can boost the effectiveness of therapies: the Mediterranean region, in this context, boasts an abundance of antioxidant-rich natural compounds [48,86].

2.1. Oxidative Stress in Cancer Development: The Role of Scavenger Enzymes

Increasing ROS release is observed during tumor development, activating mechanisms that drive tumor progression. This causes DNA damage and genetic instability, thus increasing cancer cell survival and resistance to cell death; consequently, cancer cells develop a more aggressive nature and capacity for metastasis. Additionally, high ROS levels trigger resistance mechanisms in tumor cells, which concurrently maintain protumorigenic signaling. Furthermore, oxidative stress contributes to resistance against chemotherapy [87]. Therefore, altering the oxidized/reduced state ratio may offer an oncological treatment, linked to the antioxidant effects of various natural compounds [88]. Unbalanced oxidative stress levels lead to the activation of oncogenes and reduction in the activity of tumor suppressors. High ROS production in cancer can result in the inactivation of H₂O₂ scavenger enzymes like glutathione peroxidase (GPX), glutathione reductase (GRX), catalase (CAT), superoxide dismutase (SOD), superoxide reductase, thyroxine (TRX), and peroxiredoxin (PRX) and the tumor suppressor gene phosphatase and tensin homolog (PTEN), a negative regulator of the PI3K/AKT signaling pathway, as well as the oxidation and inactivation of many other protein tyrosine phosphatases (PTPs) such as tyrosine-protein phosphatase non-receptor type 1 (PTP1B) [89]. PTEN oxidation and inactivation results in cell survival through the activation of PI3K and protein kinase B (Akt) pathways [90]. Akt is involved in cell survival through phosphorylation and the inactivation of its target proteins, including the pro-apoptotic transcription factors: forkhead box transcription factors (FOXO), BCL2 associated agonist of cell death (BAD), BCL2 associated X apoptosis regulator (BAX), and B-cell lymphoma 2 (Bcl-2)-like protein 11 (BCL2L11) [91]. Additionally, several cancers, including esophageal squamous cell carcinomas, stomach adenocarcinomas, and colorectal carcinomas (CRCs), show increased expression of genes that code for SOD proteins [92]. Tumor pathogenesis is associated with mutations in transcription factors, such as nuclear erythroid 2-related factor (Nrf2), and tumor suppressor genes, such as p53 [93]. In fact, under physiological conditions, the expression and activity of the transcription factor Nrf2 are strictly degraded by Kelch-like ECH-associated protein 1 (KEAP1), an adaptor of Cullin 3-based E3 ubiquitin ligase [94]. Cytosolic KEAP1 binding to Nrf2 triggers Nrf2 ubiquitination and proteasomal degradation; consequently, Nrf2 levels remain low under normal conditions. Oxidative stress causes the oxidation of thiol groups within KEAP1 (e.g., S-S cross-links) and this conformational

Nutrients **2025**, 17, 2354 10 of 41

change causes the release of Nrf2 in the cytosol, with its subsequent translocation to the nucleus, where Nrf2 activates antioxidant response element (ARE), which regulates antioxidant enzymes such as NAD(P)H quinone oxidoreductase (NQO1), glutathione S-transferases (GSTs), thioredoxin reductase-1 (TXNRD1), thioredoxin, ferritin, glutamate-cysteine ligase, and heme oxygenase-1 (HO-1), depending upon the binding site present in the promoter region. Nrf2 has been reported to be active and overexpressed in cancer, promoting tumor progression and cancer cell survival [95]. Its activation can also be mediated by other signal transduction pathways such as AMPK, extracellular signal-regulated kinase (ERK), c-Jun amino-terminal kinase (JNK), and PI3K/AKT [96].

2.2. Pathways Involved in Oxidative Stress-Induced Tumorigenesis

Tumorigenesis frequently involves the constitutive activation of the Nrf2 pathway, as shown in various studies. This activation arises from mechanisms including oncogene (KRAS, B-Raf proto-oncogene serine/threonine kinase (BRAF), cellular myelocytomatosis oncogene (c-myc) driven transcription, KEAP1 epigenetic suppression, or KEAP1 somatic mutations disrupting Nrf2 binding [97]. In addition, growth factors and KRAS-stimulated pathways have been shown to activate MAPK/ERK 1/2 in cancer, a protein kinase that is a member of the mitogen-activated protein kinase (MAPK) family resulting in increased cell proliferation [98]. Estrogen metabolism generates H₂O₂, which triggers the ERK1/2 signaling pathway in various cancers, such as BC; cancer cell motility and anchorage-independent growth are also impacted by the latter. Mitochondrial dysfunction-induced ROS production increase promotes NF-kB upregulation [99] which induces the up-regulation of epidermal growth factor receptor (EGFR) and its ligands through polycystin 1, transient receptor potential channel interacting (PKD1)-mediated signaling. PKD-family serine/threonine kinases act downstream of protein kinase C (PKC) and other signaling pathways, including those of Src and Abl, play a key role in detoxification from ROS and the transcription of anti-apoptotic genes [100]. The promotion of tumor progression by PKD1 involves supporting mechanisms for cancer cell survival. This includes activating ERK1/2 and regulating the JNK pathway, with effects varying based on the cellular environment [101]. Low H₂O₂ levels can trigger NF-κB, an oxidative stress detector; this can trigger the production of anti-apoptotic genes and pro-cancer cytokines like interleukin-6 (IL-6) and tumor necrosis factor alpha (TNF-α) and may activate or suppress MAPK phosphorylation. Moreover, blocking NF-kB may overcome drug resistance, a key factor in the development of chemo-resistance [102]. Reduced ROS activates hypoxia-inducible factor 1-alpha (HIF1- α), leading to the expression of genes essential for cancer growth, such as the vascular endothelial growth factor (VEGF) and its receptors; high ROS levels can directly cause oxidative DNA damage [103] and interfere with the function of epigenetic modifiers, such as DNA methyltransferases (DNMTs) and histone deacetylases (HDACs), resulting in both the hypomethylation and hypermethylation of DNA. Oxidative DNA damage causes alterations such as base pair deletions, insertions, mutations, and double-strand breaks (DSBs), forming mutagenic 8-oxo-2'-deoxyguanosine (8-oxodG), which can accumulate and cause cancer. Furthermore, telomere 8-oxo-G accumulation affects length and impairs end-capping maintenance [104]. ROS have a direct impact on the ability of transcription factors (TFs) to bind DNA: increased localization of Fos/Jun DNA-binding redox factor-1 (Ref-1) protein and ataxia-telangiectasia mutated (ATM) serine/threonine kinase—key to DNA repair—occurs in the nucleus. High ROS levels promote cancer metastasis by suppressing natural killer (NK) cell activity [105] and can regulate macrophages recruitment and cancer cell invasion. In fact, ROS contribute to the acquisition of metastatic potential by cancer cells through EMT and the expression of TF Snail, which downregulates the expression of epithelial cadherin (E-cadherin) and promotes the expression of neural

Nutrients 2025, 17, 2354 11 of 41

cadherin (N-cadherin) and vimentin. Excessive levels of ROS can lead to the activation of intrinsic and extrinsic apoptosis; in particular, mitochondrial excess ROS levels play a key role in initiating intrinsic apoptosis. High ROS levels lead to the release of cytochrome C (cyt C) into the cytosol, where it engages the apoptotic protease-activating factor 1 (APAF1), followed by the formation of the apoptosome and the activation of caspase-9 [106]. ROS can activate the extrinsic apoptotic pathway by modulating death receptors at the transmembrane level, including Fas (CD95), tumor necrosis factor receptor 1 (TNFR-1/TNF- α receptor) and tumor necrosis factor-related apoptosis-inducing ligand receptors (TRAIL-R1/2). Activated receptors, in the cytosol, recruit adaptor proteins (e.g., Fas-Associated Death Domain Protein (FADD)) and procaspases-8/10, forming death-inducing signaling complexes (DISCs) that activate caspases-8 and -10. Direct activation of effector caspases -3, -6, and -7 is possible via the latter; however, in some cases, they cleave Bid into tBid, which in turn stimulates mitochondrial outer membrane permeabilization (MOMP), blocking the anti-apoptotic activity of Bcl-2 and Bcl-XL, and leading to the release of cyt C, thereby activating the intrinsic pathway [107]. Oxidative stress and DNA damage, byproducts of chemo and radiotherapy, can cause tumor cells to develop drug resistance.

The Dual Action of Nutraceutical Compounds and Their Therapeutic Role

In vitro and *in vivo* studies support the idea that eating antioxidant-rich foods is a good way to reduce cancer risk. Some natural compounds have demonstrated dual action: they act as antioxidants to protect healthy cells from treatment-related oxidative stress, and as pro-oxidants in cancer cells, activating pro-apoptotic pathways. The antioxidant properties of bergamot polyphenolic fraction (BPF) are shown by its ability to reduce oxidative stress biomarkers and intracellular ROS levels [108]. Studies show that BPF can reduce MDA levels in rats fed a hyperlipidemic diet (HLD) [109] and in rats fed a high-fat diet (HFD) [110]. Bergamot's lipid-lowering effects, alongside reductions in MDA, Lectin-like oxidized low-density lipoprotein receptor-1 (LOX-1), and protein-kinase B (PKB), indicate improved potential, especially for those on statins [111]. BPF also improves the activity of endogenous antioxidant enzymes, including GPX and SOD. By reducing peroxynitrite, it improves antioxidant defense mechanisms, thus preventing oxidative stress-induced tissue damage. High-fat Western diet-fed (WD SW) mice treated with BPF showed reduced 3-nitrotyrosine (3-NT) in their livers compared to controls; 3-NT is a biomarker of peroxynitrite oxidative damage [112]. The protective effect of BPF on dox-induced ROS accumulation in cardiomyocytes in vivo, leading to a reduction in Dox-induced cardiomyopathy by enhancing the resident c-kit+ CD45^{neg} CD31^{neg} endogenous cardiac stem cells (eCSCs) and preventing 8-hydroxy-2'-deoxyguanosine (8-OHdG) nuclear accumulation has been demonstrated [113]. Phenolic compounds found in Cynara cardunculus extract, including caffeic acid and chlorogenic acid are effective antioxidants [114] acting as free radical scavengers and modulating the expression of genes coding for antioxidant enzymes such as SOD, GPX, and CAT [115]. Moreover, the sesquiterpene cynaropicrin has antioxidant properties [116] and pro-oxidant action in cancer cells [117]. In fact, inhibiting thioredoxin reductase (TxR), a key enzyme in cellular redox homeostasis, causes intracellular ROS increase and triggers apoptosis in cancer cells [118]. Cynaropicrin treatment increased nuclear Nrf2 in A375 human melanoma cells, as demonstrated by in vitro antioxidant assays. This is linked to a time-dependent rise in the transcription of genes that code for antioxidant enzymes like glutamate-cysteine ligase (GCL) and heme oxygenase (HMOX-1) [119]. In vivo studies reveal the effect of cynaropicrin on the antioxidant enzymes (SOD, CAT, Glutathione Peroxidase (GSH-PX)), and on 8-OHdg in the control and cerebral ischemiareperfusion (I/R) injury in rats, highlighting that it supports the antioxidants defense system against oxidative-stress [120]. Specifically, bergamot extract formulations with

Nutrients **2025**, 17, 2354

Cynara cardunculus show synergistic effects, potentially expanding treatment options for various diseases, including cancer, thanks to their antioxidant properties [121]. An *in vivo* study has shown the increasing level serum of GPX and SOD after 16 weeks of treatment with BPF and *Cynara cardunculus* phytocomplex, named Bergacyn[®], accompanied by a reduction in serum MDA levels [122].

It has been shown that oleuropein is able to increase the activity of antioxidant enzymes (i.e., SOD, GPX, GRX, CAT) and non-enzymatic antioxidant systems such as GSH [123]. It is established that oleuropein's antioxidant activity is dependent on factors such as cell type, concentration, exposure time, and oxidative stress levels [53]. In fact, its pro-oxidant action has been shown in several *in vitro* cancer cell lines, for example, MCF-7 BC cells [124], HepG2 hepatocellular carcinoma (HCC) cells [125], and HEY human epithelial ovarian cancer cell line [126], thus contributing to cell death by promoting cell damage and leading to apoptosis. Recent studies also demonstrate that oleuropein, BPF, and *Cynara cardunculus* extract lessen Dox's cardiotoxic effects within an *in vitro* rat embryonic cardiac myoblast (H9c2) model [113,127,128]. Moreover, their contribution to upholding the integrity of cellular plasma membranes' phospholipid bilayer could be pivotal via lipid content regulation. These natural compounds reportedly prevent endoplasmic reticulum (ER) dysfunction and promote apoptosis by modulating intracellular calcium [129]. Nevertheless, more research is necessary to verify the effectiveness and safety of these compounds for human use.

The preventative and therapeutic effects of quercetin and RV on various cancers are well-documented and dose-related [130]. At low doses, these agents exhibit antioxidant properties (ROS neutralization), but at higher doses, they stimulate ROS production, leading to apoptosis via the intrinsic pathway (cyt C release, caspase activation) and extrinsic pathway (death receptor activation) [131]. Low doses of quercetin and RV stimulate GSH synthesis and modulate antioxidant enzymes (SOD, CAT, GPX), offering protection. KEAP1 residue oxidation and subsequent inhibition leads to Nrf2 accumulation in the cytoplasm, followed by nuclear translocation and ARE activation [132]. Through AMPK and silent mating type information regulation 2 homolog 1 (SIRT1) activation, RV inhibits the mammalian target of rapamycin (mTOR) and deacetylates/activates forehead transcription factor O subfamily member 3a (FOXO3a), boosting antioxidant enzyme gene expression [133]. While at high doses, they interfere with mitochondrial function, generating dissipation of membrane potential, with the release of cyt C [134]. By suppressing intracellular ROS accumulation, RV protected H9c2 cardiac cells form Dox-caused toxicity [135]. Because of its strong antioxidant properties, quercetin enhances Dox's antitumor activity by inhibiting topoisomerase II (Topo II) and intercalating with DNA [136].

Several studies highlighted the biphasic effect of ferutinin [137]: low doses, in fact, show antioxidant and protective effects on healthy cells, decreasing oxidative damage. Ferutinin pretreatment at a low dose significantly reduced Dox-induced cardiotoxicity in H9c2 cells, according to Macrì et al. [138]. In addition, ferutinin at low concentrations demonstrated hyperproliferative effects with phytoestrogenic action in MCF-7 BC cells. At higher concentrations, ferutinin can increase oxidative stress in cancer cells with antiproliferative effect [139], showing a lower toxicity in healthy cells with respect to cancer cells. Bax upregulation and cyt C release, hallmarks of mitochondrial apoptosis, are dose-dependently increased by ferutinin [140]. It shows selective toxicity toward cancer cells over healthy cells, with effects varying by dose and cell type. Specifically, high ferutinin levels suppressed the growth of MCF-7 (estrogen-dependent) and MDA-MB-231 (estrogen-independent) BC cells [141]. Furthermore, ferutinin pretreatment decreased ROS and MDA buildup in lipopolysaccharide-stimulated (LPS) neurons and shielded human oligodendrocytes (MO3.13) and neurons (SH-SY5Y) from LPS-caused oxidative stress [142]. Therefore, further *in vitro* and *in vivo* research is needed to clarify the dose-dependent

Nutrients **2025**, 17, 2354 13 of 41

mechanisms of nutraceuticals' dual effects on cancer cells (Table 2) and their potential as cancer therapy adjuvants.

Table 2. Dose-dependent Action of Nutraceuticals.

Nutraceutical	Low Dose Effect (Protec- tive/Antioxidant)	High Dose Effect (Pro- Oxidant/Anticancer)	Mechanism	References
Bergamot polyphenolic fraction	↓ MDA/ROS; ↑ SOD/GPX; ↓ 3-NT/LOX-1; ↑ protective effect.	Synergistic support in redox balance, not directly cytotoxic.	↓ peroxynitrite; ↑ antioxidant enzymes (cardiomyocytes, liver).	[108–111,113,128,143]
Cynaropicrin	↑ Nrf2/SOD/CAT/GSH- PX; antioxidant in A375, brain, liver tissues.	Inhibits $TxR \rightarrow \uparrow$ ROS \rightarrow apoptosis in cancer cells.	Activates antioxidant genes (GCL, HMOX-1), pro-apoptotic in melanoma, neural, liver cells.	[117–119]
Oleuropein	↑ SOD/GPX/GRX/CAT; ↓ ROS; protects membrane and ER integrity.	↑ ROS, mitochondrial dysfunction, cyt C release; apoptotic in MCF-7, HepG2, HEY cancer cells.	Dose- and cell-type specific redox modulation.	[124,125]
Quercetin	Activates Nrf2; † GSH/SOD/GPX; inhibits Topo II; protection from Dox cardiotoxicity.	↑ROS; DNA damage; cyt C release; apoptosis via intrinsic/extrinsic pathways.	Caspase cascade, KEAP1 oxidation, ARE activation, Topo II inhibition.	[132]
Resveratrol	↑ SIRT1/AMPK/FOXO3a antioxidant genes; prevents Dox toxicity.	↑ ROS; mitochondrial a, permeability transition pore (mPTP) opening; apoptosis through caspase activation.	Mitochondrial depolarization; dual effect depending on [133–135] dose and exposure time.	
Ferutinin	Antioxidant, phytoestrogenic; ↓ ROS/MDA; protects H9c2 and neural cells.	↑ ROS, ↑ Bax, cyt C; apoptosis in MCF-7, MDA-MB-231 BC cells.	SERM-like; mitochondrial apoptosis; cell-selective effects.	[138–142]

The arrows indicate decrease (\downarrow) and increase (\uparrow), and progression (\rightarrow), respectively; MDA: malondialdehyde; ROS: reactive oxygen species; SOD: superoxide dismutase; GPX: glutathione peroxidase; 3-NT: 3-nitrotyrosine; LOX-1: lectin-like oxidized low-density lipoprotein receptor-1; Nrf2: nuclear erythroid 2-related factor; CAT: catalase; GSH-PX: glutathione peroxidase; TxR: thioredoxin reductase; GCL: glutamate-cysteine ligase; HMOX-1: heme oxygenase; GRX: glutathione reductase; Cyt C: cytochrome C; GSH: glutathione; Topo II: topoisomerase II; Dox: doxorubicin; DNA: deoxyribonucleic acid; KEAP1: kelch-like ECH-associated protein 1; ARE: antioxidant response element; SIRT1: silent mating type information regulation 2 homolog 1; AMPK: AMP-activated protein kinase; FOXO3a: forkhead transcription factor O subfamily member 3a; mPTP: mitochondrial permeability transition pore; BC: breast cancer; SERM-like: selective estrogen receptor modulator-like.

3. Highlights of the Inflammatory Process

Inflammation plays a crucial role in cancer development and metastasis, thus a key focus for cancer therapies. The growth of blood vessels, stimulated by cytokine production, aids carcinogenesis, while the resulting ROS leads to DNA damage. Moreover, the promotion of resistance mechanisms in cancer cells by inflammation can lead to chemoresistance.

Nutrients **2025**, 17, 2354 14 of 41

This occurs both through increased DNA damage due to inflammation, with activation of oncogenic bypass signaling pathways (e.g., mesenchymal–epithelial transition factor (C-MET), PI3K/AKT, MAPK), and through the action of some mediators of inflammation, including IL-6 and interleukin-1 alpha and interleukin-1 beta (IL- $1\alpha/\beta$), which modulate apoptotic and autophagic processes [144]. Tissue damage triggers inflammation—the body's complex response involving a cascade of pro- and anti-inflammatory mediators (e.g., cytokines, prostaglandins) to various insults (ischemia, trauma, infection, toxins).

The inflammatory process recruits microcirculation ECs and white blood cells, a process driven by molecular signals and balanced levels of pro- and anti-inflammatory mediators which are essential to maintain homeostasis [145]. A persistent inflammatory trigger or a failed initial response can shift an acute inflammatory to a chronic process. Aggressive cancer phenotypes, marked by increased survival, proliferation, invasion, angiogenesis, and metastasis, are often facilitated by the pro-cancer microenvironment generated from chronic inflammation.

The inflammatory response, characterized by toll-like receptor (TLR) activation, is linked to and modified by dysbiosis [146], NOD-like receptors (NLRs), C-type lectin receptors (CLRs), and retinoic acid-inducible gene (RIG)-I-like receptors (RLRs) by neutrophils/macrophages in response to pathogen-associated molecular patterns (PAMPs) [147].

Leukocyte recruitment and infiltration are mediated by chemokines like interleukin-8 (IL-8) and macrophage-inflammatory protein 2 (MIP-2) alongside inflammatory mediators including complement C5a complement fragments, platelet-activating factor (PAF), and leukotriene B4 (LTB4) [148] from the venous system to sites of tissue damage, and there is their attachment to the ECM. Activated ECs express adhesion molecules, such as the intercellular adhesion molecule 1 (ICAM-1) and intercellular adhesion molecule 2 (ICAM-2), facilitating leukocyte adhesion and diapedesis. This process involves adhesion molecules (i.e., L-P- and E-selectin) and chemokines/cytokines (i.e., TNF- α , TGF- β , IL-1 β , IL-6) secreted by activated macrophages, which regulate inflammatory cascade. The pro-inflammatory phenotypes in ECs and fibroblasts are mainly triggered by TNF- α , IL-6, and IL-1 β , which activate TNFR, interleukin-6 receptor (IL-6R), and interleukin-1 receptor (IL-1R), resulting in the activation of NF- κ B, MAPK, and Janus kinase/signal transducer and activator of transcription (JAK-STAT) signaling pathways [149].

3.1. Hallmarks of Cancer Inflammation

The research focuses on understanding the mechanisms through which inflammation impacts cancer, aiming to identify more selective therapeutic targets within specific pathways [144]. Findings from basic and clinical research demonstrate the importance of inflammatory molecules in the initiation and advancement of multiple cancer types, including breast, lung, and liver cancers [150]: these molecules could impact every phase of tumor growth, from malignant change to metastasis [151]. Conversely, acute inflammation may inhibit cancer progression: indeed, it can stimulate the immune system to help remove tumor cells. The onset of chronic inflammation increases the risk of developing highly invasive cancers. In particular, an inverse correlation between MedDiet and chronic inflammation, with lower levels of inflammatory biomarkers (i.e., CRP and IL-6) and lower risk of several cancers, has been reported [152]. Inflammation can promote tumor initiation, due to infections (e.g., Human Papillomavirus (HPV), Helicobacter pylori (H. pylori), Hepatitis B Virus (HBV), Hepatitis C virus (HCV) or carcinogens (e.g., cigarette smoke, toxic substances, ionizing radiation) [153]. Conditions such as diabetes, hyperlipidemia, and CVD are associated with low-grade inflammation that may predispose to the risk of developing cancer (e.g., BC, HCC, pancreatic cancer (PC)) [154]. The activation of Signal

Nutrients **2025**, 17, 2354 15 of 41

Transducer and Activator of Transcription proteins (STAT), especially Signal Transducer and Activator of Transcription 3 (STAT3), a transcription factor regulating genes related to cell growth, survival, and immunity, is heavily involved in cancer development across many tissues and inflammatory responses in cancers of the stomach, liver, lung, colon, and pancreas. ROS and RNS, substances produced by leukocytes and other phagocytic cells, can cause irreversible DNA damage (oncogenic mutations like RAS and myc, and the inactivation of p53 and retinoblastoma protein 1 (RB1)), increasing cancer risk. The cytokine MIF, released from macrophages, T lymphocytes, and tumor cells, can aggravate DNA damage and the accumulation of oncogenic mutations [155]. By suppressing p53's transcriptional activity, MIF interacts with it and promotes the survival of damaged cells. By suppressing pro-apoptotic gene activation and stimulating cell proliferation through PI3K/AKT and NF-кB signaling, this mechanism contributes to tumor development. The release of pro-inflammatory cytokines (TNF-α, IL-1, IL-6, IL-8, and interleukin-23 (IL-23)) from macrophages and T-lymphocytes indirectly leads to genetic instability via epithelial cell ROS production [151]. NF-kB signaling is activated via TNFR and IL-1 receptor engagement by TNF-α and IL-1 cytokines, thereby promoting a pro-inflammatory phenotype in ECs and fibroblasts. The JAK/STAT3 pathway is mainly activated by IL-6 and IL-23, contributing to the development of inflammatory tumors. In addition, the binding of IL-22 to its receptor (Interleukin-22 Receptor Subunit 1(IL-22R1)/Interleukin-10 Receptor Subunit 2 (IL-10R2)) triggers the janus kinase 1 and tyrosine kinase 2 (JAK1/TYK2) pathway, causing STAT3 phosphorylation and subsequent nuclear translocation to activate target genes involved in epithelial cell survival and regeneration. The roles of NF-KB and STAT3 pathways in tumor formation are widespread across many tissue types and strongly associated with inflammation in cancers of the stomach, colon, liver, lung, and pancreas, influencing cell proliferation, survival, angiogenesis, and immune evasion [156]. The TME comprises cancer cells, stroma, blood vessels, CAFs, ECs, pericytes, dendritic cells (DCs), and activated macrophages; these components are crucial for both antigen-specific immunity and immune tolerance, bridging innate and adaptive immunity. The TME significantly involves TAMs, which are derived from circulating monocytic precursors and are directed toward the tumor and are induced by interferon-gamma (IFN- γ)-mediated classical activation or alternatively by the helper T 2 cells (Th2) and cytokines like interleukin-4 (IL-4) and interleukin-13 (IL-13). TAMs can stimulate tumor cell proliferation, promote angiogenesis, and promote invasion and metastasis. Inflammatory-TME also includes polymorphonuclear leukocytes, and various inflammatory cells such as T lymphocytes (sometimes B cells) [157,158]. Consequently, the link between oxidative stress, chronic inflammation, and cancer development is clear. Recent findings indicate a link between the aberrant activation of Wnt\u00bb/catenin signaling and the progression of tumors and chronic inflammatory diseases. Research into Wnt/β-catenin signaling reveals opposing roles for peroxisome proliferator-activated receptor-gamma (PPARy), which is downregulated in chronic inflammation and cancer [159,160]. PPARy's role is context and tissue-specific, with evidence suggesting it can function as an oncosuppressor in certain situations, causing cell cycle arrest and apoptosis while lowering cell invasion, migration, and inflammation [161]. Deregulation in pro-inflammatory or hyperlipidic conditions promotes oncogenesis [162]. Conversely, the NF-kB pathway upregulates pro-inflammatory cytokines (interleukin-6 (IL-6), VEGF, (IL-8)), Inducible nitric oxide synthase (iNOs), TGF-β, indirectly boosts WNT/β-catenin, thus fueling inflammation and tumor growth. Survival and prognosis may be indicated by the grade of tumor infiltration, which varies by tumor type.

Nutrients **2025**, 17, 2354 16 of 41

3.2. Modulation of Inflammatory Pathways by Nutraceutical Compounds from the Mediterranean Diet

The effect of diet on acute and chronic inflammation is associated with the development of cancer. [163]. The anti-inflammatory properties of numerous natural compounds suggest their potential use as supplemental components within alternative anti-inflammatory treatment systems (Figure 2).

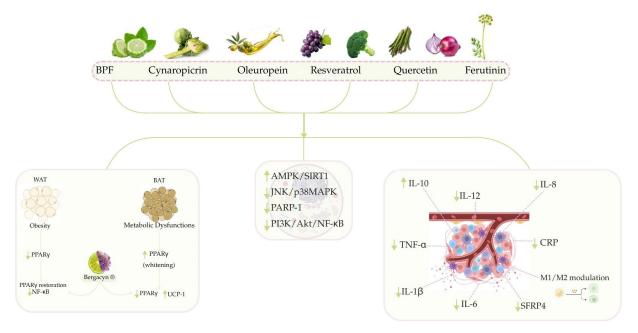


Figure 2. BPF, cyn, oleuropein, RV, quercetin, and ferutinin are able to modulate inflammatory pathways including AMPK/SIRT1, JNK/p38MAPK, PARP-1, and PI3K/Akt/NF-κB; modulate the secretome of tumor cells by reducing the secretion of pro-inflammatory cytokines (IL-12, IL-8, CRP, SFRP4, IL-6, IL-1 β , TNF- α) and increasing the secretion of anti-inflammatory cytokines (IL-10), and modulate the activation state of macrophages. Bergacyn[®] restored PPARγ levels and prevented NF-κB overexpression in obese WAT, where PPARy is significantly reduced. Whereas, in BAT, despite the increased expression of PPARγ, during metabolic dysfunction and "whitening," Bergacyn[®] reduced PPARγ. Restoration of adipose tissue homeostasis and regulation of PPARγ by Bergacyn[®] could reduce inflammation and metabolic signals involved in some cancers. The arrows indicate decrease (\$\psi\$) and increase (†), respectively. Bergamot polyphenolic fraction (BPF); cynaropicrin (cyn); resveratrol (RV); AMPactivated protein kinase/silent mating type information regulation 2 homolog 1 (AMPK/SIRT1); c-Jun amino-terminal kinase/p38 mitogen-activated protein kinase (JNK/p38MAPK); poly (ADP-ribose) polymerase-1(PARP-1); phosphatidylinositol 3-kinase/protein kinase B/Nuclear factor kappa-lightchain-enhancer of activated B cells (PI3K/Akt/NF-κB); interleukin-12, -8, -6, -1β, -10 (IL-12, -8, -6, -1β, -10); C reactive protein (CRP); secreted frizzled-related protein 4 (SFRP4); tumor necrosis factor alpha (TNF- α); peroxisome proliferator-activated receptor-gamma (PPARγ); brown adipose tissue (BAT); white adipose tissue (WAT).

3.2.1. Anti-Inflammatory Activity of Citrus Bergamia

Citrus bergamia derivatives have demonstrated anti-inflammatory activity, linked to AMPK/SIRT1 axis activation [164]. The activation of AMPK, a key regulator of energy and senescence, is linked to changes in inflammatory factors such as IL-6 and TNF- α [133]. Importantly, BPF might reduce levels of serum glucose, transaminases, gamma-glutamyl transferase (GGT), and inflammatory biomarkers such as TNF- α and CRP [165]: this prevents worsening liver inflammation and fibrosis [112]. Reduced hepatic inflammation, via IL-6 reduction and Interleukin-10 (IL-10) mRNA upregulation, has been linked to BPF supplementation [166]. By lowering phospho-JNK and phospho-p38 MAPK, it can control the release of pro-inflammatory cytokines such as IL-1 β . The reduction in inflammation and

Nutrients 2025, 17, 2354 17 of 41

hepatocellular ballooning, key to cancer initiation, also supports this effect [112,150]. This effect may also be due to BPF's antioxidant activity, which lowers excessive poly (ADP-ribose) polymerase-1 (PARP-1) activity. In disease states, the second choice has been observed to decrease the levels of the MAPK inhibitor mitogen-activated protein kinase phosphatase 1 (MKP-1), leading to elevated inflammation. Consequently, BPF's anti-inflammatory effect stems from the decreased PARP-1 repression of MKP-1, leading to JNK/p38 MAPK inhibition [167]. Findings from a randomized, double-blind, placebo-controlled study in type 2 diabetes mellitus/non-alcoholic fatty liver disease (T2DM/NAFLD) patients show that Bergacyn[®] improves NAFLD and liver fibrosis markers by modulating inflammatory biomarkers [122].

3.2.2. The Beneficial Effects of Cynaropicrin and Bergacyn[®] to Counteract Inflammation

Recently, in vivo studies evaluated Bergacyn®'s potential for tissue-specific PPARγ modulation. In obese white adipose tissue (WAT), where PPARγ is significantly reduced, Bergacyn[®] restored PPARγ levels and prevented NF-κB overexpression. Despite increased PPARy expression in brown adipose tissue (BAT) during metabolic dysfunction and "whitening," Bergacyn® downregulates PPARγ, thus promoting a thermogenic and metabolically active state [121]. PPARy's involvement in colorectal, lung, breast, prostate, and pancreatic cancer (proliferation, differentiation, and inflammation) makes the tissue-specific Bergacyn[®] modulation data highly impactful [168,169]. Moreover, the inflammatory microenvironment, a consequence of obesity-linked adipose tissue dysfunction, can promote tumor progression. Therefore, the restoration of adipose tissue homeostasis and PPARy regulation by Bergacyn[®] could lessen inflammation and metabolic signals involved in certain cancers [121] (Figure 2). Several studies highlight cynaropicrin's involvement in the inhibition of the NF-κB transcriptional activation pathway [170]. *In vitro* studies on the murine macrophage cell line RAW264.7 show its ability to inhibit inflammatory cytokine secretion following the induction of inflammation with LPS [171]. In the A375 cell line treated with cynaropicrin, the p65/RelA protein, a member of the NF-κB family, was reduced [119]. In vivo studies report that dosage-based cynaropicrin treatment on rats with cerebral I/R injury-induced significantly reversed the increased level of the pro-inflammatory cytokines (TNF- α , IL-6, and IL-1 β), and appreciably reduced the mRNA level of NF-kB [120].

3.2.3. Anti-Inflammatory Activity of Oleuropein

In vitro oleuropein has been shown to reduce the secretion of pro-inflammatory cytokines and inflammatory mediators [172,173]. In a RAW264.7 macrophage line activated with LPS (M1 phenotype), oleuropein treatment reduced Interleukin-12 (IL-12) and TNF- α levels in the cell culture supernatant and the oleuropein action on readjusting the M1/M2 macrophage polarization towards the anti-inflammatory M2 phenotype has been reported [174]. RAW264.7 cells were used to assess oleuropein's ability to reduce LPS-induced inflammation by reducing levels of IL-1 β , IL-6, and TNF- α [175,176]. Oleuropein pretreatment in LPS-activated Human Umbilical Vein Endothelial Cells (HUVECs) has been shown to potentially lower mRNA levels of pro-inflammatory cytokines like IL-1 β , IL-6, TNF- α , and IL-8 [172]. Several *in vivo* studies demonstrated oleuropein's anti-inflammatory action via reduced pro-inflammatory cytokine levels [177,178]. In addition, oleuropein treatment effectively mitigated the adverse effects of inflammation in obese mice. Specifically, the treatment led to higher PPAR γ levels in fat and the liver, and lower JNK-1 levels in the liver [179].

Nutrients **2025**, 17, 2354 18 of 41

3.2.4. The Beneficial Role of Resveratrol and Quercetin to Counteract Inflammation

Recent research indicates RV and quercetin, at varying doses, decrease inflammation by impacting SFRP4, a protein involved in obesity, diabetes, and pancreatic inflammation, thus highlighting a connection between obesity-driven inflammation and dysfunctional insulin response. Resveratrol and quercetin's potent antioxidant and anti-inflammatory effects suggest their potential as pancreatic oxidative stress drug candidates [180].

Extensive research supports the growing potential of natural compounds like resveratrol and quercetin to treat inflammation; these compounds reduced weight gain, dyslipidemia, and inflammation in a study of obese rats. Reduced body weight gain, adipocyte size, and adipose tissue mass, as well as improved serum dyslipidemia, are linked to the anti-obesity effects of quercetin and resveratrol. A quercetin and resveratrol blend's obesity-reducing effects are linked to its anti-inflammatory properties; it decreases adipokine secretion, activates AMPK α 1/SIRT1 signaling, and thus may reduce HFD-induced obesity and inflammation [181].

Further evidence supports quercetin's neuroprotective effects, potentially by promoting microglia/macrophage M2 polarization via the PI3K/Akt/NF-κB pathway. These results suggest quercetin may be a useful treatment for ischemic stroke [182].

Cardiac hypertrophy in mice was induced by implanting Ang II osmotic pumps. Researchers investigated primary neonatal rat cardiomyocytes and heart tissues to understand resveratrol's mechanism in preventing Ang II-induced cardiac hypertrophy. This research reveals important new mechanisms by which resveratrol protects against Ang II-induced cardiac hypertrophy, specifically by blocking NF- κ B signaling and pro-inflammatory cytokines. The findings add to the evidence that REV may be a promising drug for treating cardiac hypertrophy and heart failure [183].

3.2.5. Preliminary Evidence of Anti-Inflammatory Activity of Ferutinin

Antioxidant, anti-inflammatory, and anticancer effects of bioactive compounds from plants and herbs were demonstrated *in vitro* and *in vivo*.

Reducing oxidative stress lowers the risk of developing various diseases, including cancer and chronic inflammatory diseases. The antioxidant activity of flavonoids, phenolic acids, stilbenes, tannins, and lignans (phenolic plants) is demonstrated in vivo and in vitro: the presence of hydroxyl groups on their aromatic rings is responsible for reducing oxidative stress, with antioxidant capacity increasing with the number of these groups. Specifically, bergamot flavonoids have demonstrated pleiotropic effects, reducing oxidative stress and inflammation at the cellular and tissue levels. Moreover, in vitro data suggests antioxidant activity in monoterpenes, sesquiterpenes, and diterpenes, bioactive compounds extracted from various plants and fruits. In fact, sesquiterpenes show significant anticancer, anti-inflammatory, and bactericidal effects, suggesting an important role in human diseases. Different experiments suggest that sesquiterpenes offer protection in low doses, while high doses cause severe toxicity; the bipolar effect is a dose-dependent increase in the permeability of lipid bilayers and mitochondrial membranes to cations—particularly divalent cations like calcium—caused by sesquiterpenes, which are important in various pathophysiological processes [137]. Furthermore, ferutinin demonstrates anti-inflammatory effects by reducing oxidative stress. The anti-inflammatory effects of ferutinin were evaluated using an edema model. Specifically, the reduction in inflammation during the early stages was significant, likely due to histamine and serotonin. Notably, at this stage, ferutinin demonstrated a significantly more potent anti-inflammatory effect compared to the aspirin control dose. This study implies ferutinin's anti-inflammatory action likely results from blocking histamine and/or serotonin activity [184]; however, further studies are needed to better explain the pathway under these mechanisms.

Nutrients **2025**, 17, 2354 19 of 41

4. Cell Cycle Regulation: Key Mechanisms and Links to Tumorigenesis

A misregulation in cell cycle regulation can promote the development of diseases, including cancer. The cell cycle is controlled by molecular mechanisms involving distinct CDK-cyclin complexes, arising from the interplay between cyclin-dependent kinases (CDKs) and cyclins [185], and through three checkpoints: (i) G1 checkpoint, (ii) G2 checkpoint, and (iii) M checkpoint, also known as spindle assembly checkpoint (SAC). The G1 checkpoint's activity is determined by growth factors, capable of both promoting and suppressing cell growth, thus monitoring the transition between the G1 phase and the S phase. The regulation of the G1 checkpoint hinges on the crucial roles played by CDKs and tumor suppressor proteins, notably p53 and pRb. If a cell identifies damage or irregularities, it activates DNA repair; if this fails, apoptosis is triggered.

The tumor suppressor p53 controls cell metabolism, redox balance, and DNA repair, inducing cell cycle arrest and cell death. Cell cycle arrest results from p53 binding to upstream promoter regions of the p21 gene, leading to its transcriptional activation; in particular, p21 protein inhibits cyclin E/Cdk2 and cyclin D/Cdk4/6 complexes, resulting in cell cycle arrest in the G1 phase [186]. Findings suggest that zinc finger protein 57 (ZFP57), a key regulator of BRCA1 expression in ovarian cancer, might impact the G1 cell cycle checkpoint [187]. The silencing of CEP192, a novel gene involved in the development of NAFLD and HCC, has been recently shown to cause cell cycle arrest in the G0/G1 phase of cancer cells, thereby decreasing their proliferation and self-renewal. These results reveal the onco-immunological involvement of CEP192 in the establishment of a TME [188]. The cell uses the G2 checkpoint to confirm successful DNA replication before beginning mitosis. Its regulation involves a complex network including CDKs and checkpoint kinases (Chk1 and Chk2), along with other proteins and signaling pathways. Overexpression of polo-like kinase 1 (PLK1), a crucial mitotic checkpoint protein for cell cycle progression, is associated with unfavorable prognoses in some cancers (e.g., pancreatic adenocarcinoma) [189]. The M checkpoint monitors kinetochore-microtubule binding during mitosis; its regulation involves Mad, Bub proteins, CDKs, and other signaling pathways [190]. The regulation of all cell cycle phases, including G1/S and G2/M transitions, by the PTEN oncosuppressor gene is crucial for cell proliferation and survival. Furthermore, it plays a role in preserving chromosome structure, safeguarding the genome against structural and numerical chromosomal instability (CIN). CDKs, specifically A, B, D, and E, are key to the molecular mechanisms governing cell cycle control. Cyclins, each produced and degraded at specific cell cycle stages, bind to particular CDKs, forming active complexes essential for each phase and their activity is positively modulated by cyclins and inhibited by CDK inhibitors (CKIs). Transcriptional and post-translational modifications, along with the ubiquitin-proteasome system's rapid degradation of cyclins and CKIs, control the activation state. Dysregulation of cyclin expression and activity, and mutations or changes in the genes that regulate checkpoints, can lead to uncontrolled cell division and contribute to cancer development [191].

4.1. Mechanistic Insights into Cell Cycle Alterations in Cancer Development

Cell cycle dysregulation arises from multiple pathways: checkpoint failures, spindle checkpoint defects, CDK/cyclin mutations, abnormal cyclin D and E levels, and telomerase-driven immortality. The initial step in tumor formation is cell immortalization, or the evasion of senescence; the immortalization is related to the activity of enzyme telomerase, which increases telomere length at the ends of chromosomes [192]. Common alterations in the cell cycle include elevated levels of CDK4/6 and CDK2, both vital kinases in the G1 to S phase progression. Phosphorylated Rb underlies these alterations, leading to E2F release and activation of genes essential for cell cycle's S phase initiation. Overactive complexes

Nutrients 2025, 17, 2354 20 of 41

correlate with boosted cancer cell growth, aging, migration, and blood vessel formation, alongside apoptosis resistance and therapy resistance (like against CDK inhibitors) in cancers such as breast and ovarian cancers [193]. Genes regulating cell cycle checkpoints, including p53 and FRP1, also show mutations or deletions in cancer cells. The lack of p53 crucially affects cell cycle checkpoint regulators, leading to the increased expression of genes such as p21 (a growth inhibitor) and pro-apoptotic genes, mainly after DNA damage halts the cell cycle at G1 [186]. Uncontrolled cell cycle progression may result from the loss of oncosuppressor genes, including Rb1, p16INK4A, p15INK4B, and p14ARF. Furthermore, mutations in p16, Rb loss, or increased CDK2 activity can lessen the effectiveness of CDK4/6 inhibitors (e.g., palbociclib, ribociclib, abemaciclib). This has prompted the development of combination, or second-generation, strategies to overcome therapeutic resistance [193]. The link between checkpoint failure and cancerous phenotypes in normal cells is established; however, the specific genetic alterations responsible are still unknown. Furthermore, the direct causal link between checkpoint malfunction and cell transformation, versus the involvement of other regulatory genes after immortalization, requires further investigation. In cancer treatment, genes involved in checkpoints may be useful as diagnostic tools and as targets for new drugs [194].

4.2. Nutraceutical-Induced Cell Cycle Modulation in Cancer and Chemotherapy—Induced Cytotoxicity

The effects of natural compounds on inhibiting tumor cell proliferation by modulating specific cell cycle checkpoints and interfering with mitotic progression have been shown; their effects on cell cycle regulation are reported in Table 3. This suggests both their potential use as coadjuvants in the treatment of some types of cancer, and as a treatment to reduce the toxic effects induced by chemotherapeutic drugs in non-cancerous cells [195] (Figure 3).

Natural Compound/Plant	Cell Cycle Phase	Molecular Mechanism	Cell Type	References
Citrus bergamia	G1/S	Silent mating type information regulation 2 homolog 2 (SIRT2)/Akt/p53 ↑ apoptosis	THP-1, SH-SY5Y	[196–198]
Cynaropicrin	G2/M	↓ Pyruvate kinase M2 (PKM2)/PARP	CAL-62, 8505C, SW1736, A549	[199,200]
Oleuropein	G1/S	↑ p21, p53, CKIs	MCF-7, MDA-MB-231, MDA-MB-468	[201–203]
Resveratrol	S	↑ p53/p21/p27/Bax/p53 upregulated modulator of apoptosis (PUMA) ↓ cyclins D/E ↓ cyclin-dependent kinase 2, 4, 6 (CDK2/4/6)	C-3, LNCaP, A431, T47D	[204–208]
Quercetin	G1/G2/M			[209–212]
Ferutinin	↓G1 and restoring cell cycle	↑ apoptosis ↑ protective effect	H9c2	[138,140,213]

Table 3. Effects of Natural Compounds on Cell Cycle Regulation.

The arrows indicate decrease (\downarrow) and increase (\uparrow), respectively. SIRT2: silent mating type information regulation 2 homolog 2; PKM2: pyruvate kinase M2; PARP: poly-ADP-ribose-polymerase; p53: tumor protein p53; CKIs: CDK inhibitors; Bax: BCL2 associated X, apoptosis regulator; PUMA: p53 upregulated modulator of apoptosis; CDK: cyclin-dependent kinase; Rb: retinoblastoma protein.

The anti-proliferative effects of *Citrus bergamia* natural derivatives on different cancer cell types were evaluated through cell cycle arrest in multiple *in vitro* studies [143]. Human leukemia THP-1 cells treated with Flavonoid-Rich Extract (FRE) of BJ showed cell cycle arrest in the S phase, leading to apoptosis through the SIRT2/AKT/p53 pathway [196];

Nutrients 2025, 17, 2354 21 of 41

in addition, *Citrus bergamia* juice decreased human neuroblastoma SH-SY5Y cell growth *in vitro* via G1 cell cycle arrest and reduced cell adhesion [197]. Animal studies showed that FRE of BJ decreased levels of baculoviral inhibitor of apoptosis repeat-containing 5 (Birc5) and the cell cycle regulator p21, potentially increasing tumor cell death [198].

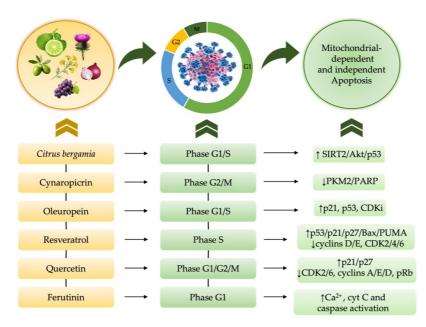


Figure 3. The anti-proliferative effects of natural extract derivatives on different cancer cell types were evaluated through cell cycle arrest in multiple in vitro studies. Citrus bergamia has been shown to possess an antiproliferative effect by arresting the cell cycle in the G1/S phase through the modulation of the SIRT2/Akt/p53 pathway. Cynaropicrin arrested the cell cycle in the G2/M phase by modulating the PKM2/PARP pathway. Oleuropein exerts an antiproliferative effect by arresting the cell cycle in the G1/S phase by increasing p21/p53/CDKi. RV treatment arrested the cell cycle in the S phase by modulating p53/p21/p27/Bax/PUMA and the reduction in cyclins D/E and CDK 2/4/6. Treatment with quercetin causes cell cycle arrest in G1/G2/M phases with the modulation of p21/p27 and reduction in CDK 2/6 and cyclins A/E/D and phosphorylation of Rb. Ferutinin treatment has been shown to exsert an antiproliferative effect leading to cell cycle arrest in the G1 phase by increasing intracytoplasmic Ca 2+ concentration, inducing cyt C release and caspases activation. The arrows indicate decrease (\downarrow) and increase (\uparrow), respectively. Silent mating type information regulation 2 homolog 2 (SIRT2); protein kinase B (Akt); tumor protein p53 (p53); Cynaropicrin (Cyn); Pyruvate kinase M2 (PKM2); poly-ADP-ribose-polymerase (PARP); tumor protein p21/p53; Cyclin-dependent kinase inhibitor (CDKi); Resveratrol (RV); p53/p21/p27; BCL2 associated X, apoptosis regulator (Bax); p53 upregulated modulator of apoptosis (PUMA); Cyclin-dependent kinases (CDK 2/4/6); Cytochrome C (Cyt C).

Other evidence highlighted that cynaropicrin could inhibit cellular growth by cell cycle arrest, mainly in the G2/M phase, in thyroid cancer cell lines CAL-62, 8505C, and SW1736 cells, by a dose- and time-dependent way [199]. This also demonstrates PKM2 inhibition in A549 cells, causing increased p53, decreased DNA repair enzyme PARP, with a subsequent cell cycle arrest [200].

Oleuropein significantly modulates the cell cycle, notably by upregulating CDK inhibitor levels, thus causing cell cycle arrest, and by increasing p53 and p21 levels, which then affects gene expression to activate both intrinsic and extrinsic apoptosis pathways [201]. *In vitro* investigations using MCF-7 and MDA-MB-231 BC cells revealed that oleuropein treatment induced cell cycle arrest at the G1 and S phases [202]; its effects on MDA-MB-231 and MDA-MB-468 BC cell lines include cell proliferation inhibition, apoptosis induction through S-phase arrest, and caspase-1, -4, and -14 expression [203].

Nutrients 2025, 17, 2354 22 of 41

Several *in vitro* studies [209] indicate quercetin's role in modulating the cell cycle. Myeloid cell line p39 and osteosarcoma cells HOS showed increased G1 cell cycle arrest after treatment, accompanied by decreased CDK-2, -6, cyclins A, D, E, Rb phosphorylation, and increased p21 and p27 expression [210]. Furthermore, quercetin triggered G1 cell cycle arrest in oral squamous cell carcinoma cell lines YD10B and YD38 OSCC [211].

It has been suggested that quercetin treatment leads to G2/M cell cycle arrest in T47D and KON BC cells and A375 melanoma cells [212].

Moreover, RV's *in vitro* effects included enhanced G1 to S phase transition, S phase cell cycle arrest, and reduced cancer cell proliferation and viability. Prostate cancer cells (LNCaP and PC-3) treated with RV showed reduced expression of cyclins D1 and E, CDK4, and cyclin D1/CDK4 kinase activity [204]. RV's ability to decrease A431 and colon cancer cell proliferation has been linked to lower levels of cyclins D1/D2/E2, CDK2/4/6, and higher levels of p21 and p27 [205]. RV was found to activate p53 along with its target genes p21, p27, Bax, PUMA, MDM2, and cyclin G [206]. Therefore, RV may stimulate the G1-to-S phase transition in BC cells, subsequently halting their progression through the S phase, thus diminishing their proliferation and viability [207]. In addition, it has been reported that RV reduces *Versican* (VCAN) secretion by CAFs, presumably by modulating the cell cycle [208].

Exposure to ferutinin leads to a decreased G1 phase and increased cell death, modifying the cell cycle profile [140]. In addition, it was shown that ferutinin, thanks to its cell-type dependent activity, restores the cell cycle in H9c2 myoblasts incubated with Dox [138], suggesting its protective role in cell cycle progression, highlighting its potential as a therapeutic agent in cancer treatment and in mitigating chemotherapy-induced cardiac damage (CIC) (Figure 4) [213].

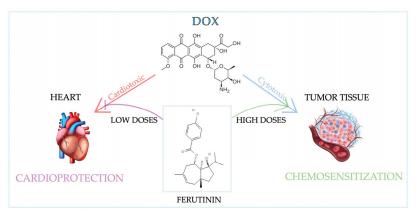


Figure 4. The tissue-dependent and dose-dependent effect of ferutinin. At low doses, ferutinin ca counteract the cardiotoxic effect of Dox. At high doses, ferutinin can enhance Dox chemosensitization in tumor tissues. Doxorubicin (Dox).

5. The Human Microbial Landscape in Cancer Development

The human microbiota comprises approximately 300 bacterial, fungal, and viral species, totaling almost 40 trillion microorganisms [214,215]. Over 97% of the human microbiota is located in the gastrointestinal tract, mainly the colon, and is known as the gut microbiota. In addition, current findings demonstrate a diverse microbiome within previously considered sterile tumor tissues (lung, breast, liver), which is closely associated with oncogenesis [216,217]. Consequently, the intratumoral microbiota (ITM) concept emerged and ITM's anti- and pro-tumorigenic effects have been demonstrated through mechanistic studies [218]. Tumor-inhibiting metabolites are produced by T and NK cells activated by the ITM. These may raise oxidative stress, causing genomic instability and mutations, affecting epigenetics, weakening immunity, and stimulating inflammation, thus, leading to

Nutrients 2025, 17, 2354 23 of 41

tumor initiation and progression [219], representing an integral part of TME [218]. Tumors in mucosal organs, including the colon, cervix, esophagus, and lung, are colonized by the ITM [220].

Non-mucosal organs such as the pancreas, liver, breast, and prostate also contained ITM, which suggests the existence of both adjacent tissue invasion and hematogenous spread during colonization [218]. This supports the hypothesis that TME colonization by the ITM may result from malformed blood vessels [217]. Tumor subtype differences in intratumoral microbial composition have been demonstrated [221]. Tumors can arise from viral infections and persistent bacterial inflammation, such as that caused by *H. pylori* and gastritis.

Evidence on the colon cancer microbiota has shown the presence of bacteria *Bacteroides* fragilis, Escherichia coli (E. coli), and Fusobacterium nucleatum—frequently detected in malignant pancreatic and BC—and fungal species such as Candida albicans [219]. The species of Blastomyces are prevalent in lung cancers, while the species of Malassezia are abundant within BC [216]. TME regulation originates from both the tumor and gut microbiota, impacting immune responses and cancer cell metabolism; consequently, this crosstalk between intratumoral and gut microbiota might elicit specific immune reactions [222]. Metagenomics, using next-generation sequencing (NGS) and computational analysis of 16S rRNA amplicons, has enabled the characterization of gut microbiota diversity and abundance [223,224]. Firmicutes and Bacteroidetes are the most prevalent phyla among the roughly 1000 bacterial species in the gut microbiota. The gut's microbiome plays a critical role in numerous physiological processes, including vitamin production, the metabolism of food components, protection from pathogens, and the shaping of the host's immune system [225]. The pharmacomicrobiomic concept has established the gut microbiota's role in influencing drug effects, such as chemotherapy. In fact, the gut's microbes can alter drug metabolism, impacting their absorption, effectiveness, or safety [226]. Dysbiosis, an imbalance of gut microbiota, is linked to cardiovascular, metabolic, and neurodegenerative diseases, as well as cancer [227]. Studies of the gut microbiome via metabolomics and metagenomics show its dual impact on cancer prevention, onset, and therapy, having both suppressing and oncogenic activity [228]. Several studies indicate that communication between colorectal cells and gut bacteria is vital for the body's overall health and immune response, especially in metabolism. The influence of gut microbiota on cancer initiation, progression, and metastasis, as well as its potential in cancer treatment, is a promising area in precision medicine [229].

Emerging Connections and Therapeutic Perspectives of Gut Microbiota and Nutraceuticals in Cancer

An interplay between the gut microbiota, diet-induced inflammation, and cancer pathogenesis has been found. In fact, an altered microbiota associated with chronic inflammation in colon, pancreatic, breast, gastric, liver, and prostate cancer, has been identified. Consequently, investigations on nutraceutical compounds potentially able to modulate the microbiota, with implications on cancer pathogenesis, are crucial. A correlation between diet and healthy gut microbiota has been demonstrated, highlighting the implication of dietary natural compounds in affecting cancer through the gut microbiota [230]; thus, this relationship between diet and cancer through the gut microbiota may provide new insights for cancer treatment [231] (Figure 5). Specifically, metabolic changes like obesity are linked to persistent low-level inflammation, a connection between obesity and cancer supported by many studies. The increasing prevalence of excessive fat dietary intake is linked to the adoption of HFD in many countries, with a lack of fiber intake [232]. In comparison to traditional rural populations, HFD populations showed reduced bacterial diversity [233]. HFDs impact gut microbiota to produce more leucine, subsequently activating the mam-

Nutrients 2025, 17, 2354 24 of 41

malian target of rapamycin complex 1 (mTORC1) in bone marrow, leading to increased polymorphonuclear myeloid-derived suppressor cells (PMN-MDSC) differentiation and faster BC growth through the gut–bone marrow–tumor axis.

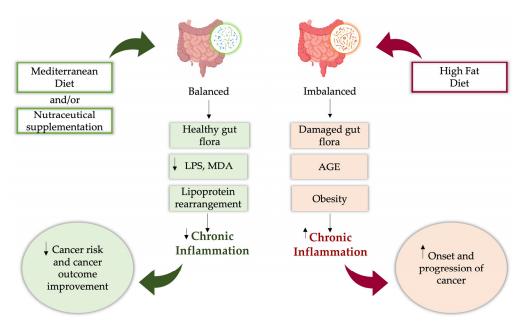


Figure 5. Mediterranean-style dietary model and/or nutraceutical supplementation are directly related to health gut microbiota. The MedDiet, rich in fiber and natural compounds such as polyphenols, supports a balanced microbiota, reduces inflammation, LPS and MDA levels, promotes lipoprotein rearrangement, and contributes to reduction in cancer risk and cancer outcome improvement. Controversially an HFD alters the composition of the microbiota, promoting AGEs production, with increased chronic inflammatory state—associated with obesity, and activating the gut–bone marrow–tumor axis, resulting in increased onset and progression of cancer. The arrows indicate decrease (\pmatheta) and increase (\epsilon), respectively. Mediterranean Diet (MedDiet); lipopolysaccharide (LPS); malondialdehyde (MDA); high fat diet (HFD); advanced glycation end product (AGE).

Evidence indicates a positive correlation between good adherence to the MedDiet and the health of the gut microbiota and could potentially represent an important pattern in reducing the incidence of a range of diseases, including cancer [234]. Several natural compounds hold promise as gut microbiota modifiers and cancer prevention/treatment aids. Notably, many dietary polyphenols can beneficially alter the gut microbiota and directly act as antioxidants, synergistically enhancing probiotic effects [235]. Polyphenols such as RV and quercetin boost intestinal health by modulating Nrf2 and NF-kB, thus improving barrier function, repairing the gut lining, reducing inflammation, and regulating immunity. RV and quercetin modulate intestinal tight junction protein expression and distribution, thus raising trans-epithelial electrical resistance (TEER), according to reports. Therefore, they beneficially impact the gut barrier, helping to manage numerous diseases stemming from gut barrier dysfunction [236]. In diet-induced obese mice, oleuropein's protection of metabolism and vasculature is linked to its immunomodulatory effects and improved gut barrier function by reducing dysbiosis [179]. A recent in vivo study highlighted that a bergamot polyphenol extract (BPE) with micronized albedo and pulp fibers (BMF) helps maintain gut microbiota health in animals fed an HFD. A four-week trial using a 50/50% BPE/BMF mixture in rats fed a HFD resulted in significantly better metabolic profiles and lower gut LPS levels, which are vital in cancer development [237,238], reducing MDA levels and leading lipoprotein size re-arrangement. This effect is related to improvements in gut microbiota composition, particularly through the modulation of the Gram-negative bacteria Proteobacteria and the dominant phyla Firmicutes and Bacteroidetes [110]. The findNutrients 2025, 17, 2354 25 of 41

ings suggest that the interaction between nutraceuticals, microbiota, and cancer is a highly promising field of research, and could offer new strategies for prevention and therapeutic support in oncology. Indeed, a complex interplay of various cell types with diverse metabolic states and shared metabolites creates the TME. The complex cellular structure produces a profile of available nutrients and reactive metabolites, including advanced glycation end-products. The literature analysis reveals that the receptor for advanced glycation end products (RAGE) and its ligands link energy metabolism (often disrupted by mitochondrial dysfunction) to immune responses (shaped by local microbiota), impacting tumor development. Investigating metabolic processes in cancer and immune cells reveals novel therapeutic strategies, potentially broadening immunotherapy's reach and effectiveness. Targeting the RAGE and its ligands in immunotherapy shows promise for treating CRC, according to the literature review [239].

6. Limitations of Nutraceutical Supplementation: Current Challenges and Emerging Evidence-Based Strategies

Although nutraceuticals offer proven benefits, their widespread use in clinical practice is hindered by considerable limitations. Factors such as the poor bioavailability of active compounds, inconsistent dose-response, and insufficient clinical trial data hinder progress [240]. In addition, responses vary considerably between individuals due to genetic, gut microbiome, dietary, age-related, and health factors, complicating the determination of ideal dosages [241–243]. While exhibiting significant *in vitro* potential, the bioavailability of polyphenols (frequently researched for nutraceutical purposes) is low due to digestive breakdown and limited absorption in the upper gut. Primarily, only a small percentage of low molecular weight polyphenols (like glycosylated flavonoids) are absorbed in the upper small intestine, mainly the duodenum and jejunum. Most complex polyphenols bypass absorption until reaching the colon, undergoing microbial biotransformation into absorbable simpler metabolites (e.g., phenolic acids) [244]. In addition, a recent study demonstrated that innovative natural formulations significantly increased plasma metabolite levels in treated animals, overcoming the poor bioavailability of polyphenols and maximizing their beneficial effects [46]. Nutraceutical safety and efficacy, and their contribution to health, will improve if "bed-to-bench" issues are addressed.

7. Conclusions

This review summarizes results and evidence showing that Mediterranean nutraceuticals hold promise in cancer prevention and treatment. The antioxidant, anti-inflammatory, and pro-apoptotic properties of Mediterranean nutraceuticals, such as polyphenols and phytochemicals from Citrus bergamia Risso & Poiteau, Cynara cardunculus, Olea europea L., quercetin, resveratrol, and Ferula communis L., are promising as complementary treatments against cancer, providing a solid basis for considering them as effective adjuvants in conventional cancer therapies. Crucially, many nutraceuticals demonstrate dual dose-dependent effects; low concentrations act as antioxidants, and high concentrations induce selective cancer cell death via oxidative stress. This two-stage process holds significant promise for preventing and enhancing the effectiveness of cancer treatment: better outcomes and reduced side effects, particularly cardiotoxicity, may result from using them with standard cancer treatments. Furthermore, the interaction of natural compounds with the main molecular pathways involved in chronic inflammation (e.g., NF-κB, STAT3, PI3K/Akt) and oxidative stress (e.g., Nrf2/KEAP1, AMPK/SIRT1) highlights their potential in remodeling the TME. The active control of metastasis and chronic inflammation is suggested by the ability to adjust macrophage polarization, cytokine production, and the innate immune response. Developing validated anticancer therapies critically depends on integrating

Nutrients 2025, 17, 2354 26 of 41

nutraceuticals, oncology, and pharmacology to optimize drug mechanisms, combinations, nutraceutical dosages (BPF, cynaropicrin, quercetin, RV, and ferutinin), and therapeutic translation. Despite this, natural compounds are not intended to replace conventional cancer treatments like chemotherapy, but rather to enhance them in a personalized approach for each patient. Low bioavailability, lack of extensive clinical trials, and inconsistent patient responses are delaying the clinical translation of these results. The efficacy of nutraceuticals in oncology requires confirmation through rigorous randomized controlled trials to implement positive preclinical results into proven therapeutic applications. Further *in vitro* and *in vivo* studies are needed to optimize bioavailability (e.g., formulation with nanocarriers), characterize pharmacokinetics, determine optimal dosages, and evaluate long-term safety. In closing, the emerging concept of onconutraceuticals represents an integrated paradigm that combines prevention, therapeutic support, and personalized medicine. In this context, the "Mediterranean" approach could offer a sustainable and evidence-based dietary-pharmacological model, useful not only for the management of neoplastic disease but also for the promotion of global health.

Author Contributions: Conceptualization, V.M., R.M. and C.A.; writing—original draft, C.A., R.M. and M.S.; writing—review and editing, G.R., S.U., C.A. and J.M.; Tables and graphs, G.R., S.U. and C.A.; supervision, V.M., C.M. and E.P.; funding acquisition, V.M. and C.M. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by PON-MIUR 03PE000_78_1, PON-MIUR 03PE000_78_2, and PRIR Calabria Asse 1/Azione 1.5.1/FESR (Progetto AgrInfra Calabria), SAP01-2023-000077—Bando A Cascata Rome Technopole Spoke 1 e Spoke 6-PRODIGI (CUP B83C22002820006).

Conflicts of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as potential conflicts of interest.

Abbreviations

8-oxodG	8-oxo-2'-deoxyguanosine
8-OHdG	8-hydroxy-2'-deoxyguanosine

3-NT 3-nitrotyrosine

AGEs advanced glycation end products

Akt protein kinase B

AMPK AMP-activated protein kinase
APAF1 apoptotic protease-activating factor 1
ARE antioxidant response element
ATM ataxia-telangiectasia mutated
ATP Adenosine Triphosphate

BAD BCL2 associated agonist of cell death

BAT brown adipose tissue

Bax BCL2 Associated X, Apoptosis Regulator

BC breast cancer

BCL2L11 B-cell lymphoma 2 (Bcl-2)-like protein 11

BEO bergamot essential oil

Birc5 Baculoviral Inhibitor of Apoptosis Repeat-containing 5

BMF albedo and pulp-derived micronized fibers

BJ bergamot juice

BPE bergamot polyphenol extract
BPF bergamot polyphenolic fraction

BRAF B-Raf proto-oncogene serine/threonine kinase c-Met mesenchymal-epithelial transition factor c-myc cellular myelocytomatosis oncogene Nutrients **2025**, 17, 2354 27 of 41

CAFs cancer-associated fibroblasts

CAT catalase

CDK cyclin-dependent kinase
CDKs cyclin-dependent kinases
Chk1 checkpoint kinases 1
Chk2 checkpoint kinases 2

CIC chemotherapy-induced cardiac damage

CIN chromosomal instability

CKIs CDK inhibitors

CLRs C-type lectin receptors
COX-1 cyclooxygenases 1
COX-2 cyclooxygenases 2
CRC colorectal carcinoma
CRP C reactive protein
CSCs cancer stem cells
CVD cardiovascular disease

Cyn cynaropicrin
Cyt C cytochrome C
Daun daunorubicin
DCs dendritic cells

DISCs death-inducing signaling complexes

DM diabetes mellitus
DNA DeoxyriboNucleic Acid
DNMTs DNA methyltransferases
DSBs double-strand breaks

Dox doxorubicin
E. coli Escherichia coli
EC endometrial cancer
ECM extracellular matrix
ECs endothelial cells

eCSCs endogenous cardiac stem cells
EFSA The European Food Safety Authority
EGFR epidermal growth factor receptor
EMT Epithelial–Mesenchymal Transition
eNOS endothelial nitric oxide synthase

ER endoplasmic reticulum

ERK extracellular signal-regulated kinase ERK1/2 extracellular signal-regulated kinase ½

 $\begin{array}{ll} ER\alpha & Estrogen\ receptor\ alpha \\ ER\beta & Estrogen\ receptor\ beta \\ EVOO & extra\ virgin\ olive\ oil \end{array}$

FADD Fas-Associated Death Domain Protein FOXO Forkhead box transcription factors

FOXO3a Forkhead Transcription Factor O Subfamily Member 3a

FRE Flavonoid-Rich Extract
GCL glutamate-cysteine ligase
GGT gamma-glutamyl transferase
GPX glutathione peroxidase
GRX glutathione reductase
GSH-PX Glutathione Peroxidase

GSH glutathione

GSK-3β glycogen synthase kinase 3 beta

GST glutathione transferase GSTs glutathione S-transferases Nutrients 2025, 17, 2354 28 of 41

 $\begin{array}{ll} \text{H. pylori} & \text{Helicobacter pylori} \\ \text{H}_2\text{O}_2 & \text{Hydrogen Peroxide} \end{array}$

H9c2 rat embryonic cardiac myoblast

HBV Hepatitis B Virus

HCC hepatocellular carcinoma

HCV Hepatitis C virusHDACs histone deacetylasesHDL high density lipoprotein

HFD high-fat diets

HIF1- α hypoxia-inducible factor 1-alpha

HLD hyperlipidemic diet

HMOX-1 heme oxygenase

HO-1 heme oxygenase-1

HOCl hypochlorous acid

HPV Human Papillomavirus

HRAS Harvey Rat sarcoma virus

HRT hormone therapy

HUVECs Human Umbilical Vein Endothelial Cells

I/R ischemia-reperfusion

IARC International Agency for Research on Cancer

ICAM-1 intercellular adhesion molecule 1 ICAM-2 intercellular adhesion molecule 2

IFN-γ interferon-gamma IL-10 Interleukin-10

IL-10R2 Interleukin-10 Receptor Subunit 2

IL-12 Interleukin-12 IL-13 interleukin-13

 $\begin{array}{ll} IL\text{-}1R & \text{interleukin-1 receptor} \\ IL\text{-}1\alpha & \text{Interleukin-1 alpha} \\ IL\text{-}1\beta & \text{Interleukin-1 beta} \end{array}$

IL-22R1 Interleukin-22 Receptor Subunit 1

IL-23 interleukin-23IL-4 interleukin-4IL-6 interleukin-6

IL-6R interleukin-6 receptor

IL-8 interleukin-8

iNOs Inducible nitric oxide synthase ITM intratumoral microbiota

JAK-STAT Janus kinase (JAK)/signal transducer and activator of transcription (STAT)

JAK1/TYK2 Janus kinase 1 and Tyrosine kinase 2

JNK-1 c-Jun N-terminal kinase 1 JNK c-Jun amino-terminal kinase

KEAP1 Kelch-like ECH-associated protein 1

KRAS Kirsten rat sarcoma viral oncogene homolog

LDL low density lipoprotein

LOX-1 Lectin-like oxidized low-density lipoprotein receptor-1

LOX lipoxygenase LPS lipopolysaccharide LTB4 leukotriene B4

MAPK Mitogen-Activated Protein Kinase

MDA Malondialdehyde

MDSCs myeloid-derived suppressor cells

MedDiet Mediterranean Diet

MIP-2 Macrophage-inflammatory protein 2

Nutrients 2025, 17, 2354 29 of 41

MKP-1 Mitogen-activated protein kinase phosphatase 1 MOMP mitochondrial outer membrane permeabilization

MS metabolic syndrome

mTOR mammalian target of rapamycin

mTORC1 mammalian target of rapamycin complex 1

MUFAs monounsaturated fatty acids

NADPH nicotinamide adenine dinucleotide phosphate

NAFLD Non-alcoholic fatty liver disease

NF-κβ Nuclear factor kappa-light-chain-enhancer of activated B cells

NGS next-generation sequencing

NK natural killer

NLRs NOD-like receptors

NOTCH Notch signaling pathway

NQO1 NAD(P)H quinone oxidoreductase 1

NRAS Neuroblastoma RAS viral oncogene homolog

Nrf2 nuclear erythroid 2-related factor

O₂ superoxide anion

O₂ dioxygen

OMWW Olive Mill Wastewater

OxLDL oxidized low-density lipoprotein

P53 tumor protein p53 PAF platelet-activating factor

PAMPs pathogen-associated molecular patterns

PARP-1 poly (ADP-ribose) polymerase-1. PARP poly-ADP-ribose-polymerase

PC pancreatic cancer

PI3K phosphatidylinositol 3-kinase

PKB protein-kinase B PKC protein kinase C

PKD1 Polycystin 1, Transient Receptor Potential Channel Interacting

PKM2 Pyruvate kinase M2 PLA2 phospholipase A2 PLK1 polo-like kinase 1

PMN-MDSC polymorphonuclear myeloid-derived suppressor cells PPARγ peroxisome proliferator-activated receptor-gamma

PRX peroxiredoxin

PTEN Phosphatase and tensin homolog PTMs post-translational modifications

PTP1B Tyrosine-protein phosphatase non-receptor type 1

PTPs Protein Tyrosine Phosphatases PUFAs polyunsaturated fatty acids

PUMA p53 upregulated modulator of apoptosis RAGE Receptor for advanced glycation end products

RAS Rat sarcoma virus

Rb Retinoblastoma protein

Ref-1 Redox Effector Factor-1

RLRs RIG-I-like receptors

RNS Reactive nitrogen species

ROS Reactive oxygen species

RV Resveratrol

SAC spindle assembly checkpoint

SERM selective estrogen receptor modulator

SIRT1 silent mating type information regulation 2 homolog 1 SIRT2 Silent mating type information regulation 2 homolog 2 Nutrients 2025, 17, 2354 30 of 41

SFRP4 secreted frizzled-related protein 4

SOD superoxide dismutase

STAT Signal Transducer and Activator of Transcription
STAT3 Signal Transducer and Activator of Transcription 3

T2DM type 2 diabetes mellitus

TAMs tumor-associated macrophages
TEER trans-epithelial electrical resistance

TFs transcription factors

TGF-β Transforming growth factor beta

Th2 helper T 2 cells TLR Toll-like receptor

TME tumor microenvironment TNF- α Tumor Necrosis Factor Alpha TNFR-1 tumor necrosis factor receptor 1

Topo II topoisomerase II

TRAIL-R1/2 tumor necrosis factor-related apoptosis-inducing ligand receptors

Tregs regulatory T-cells

TRX thyroxine

TXNRD1 thioredoxin reductase-1 TxR thioredoxin reductase UV Ultraviolet radiation

VEGF Vascular Endothelial Growth Factor

VOO virgin olive oil
WAT white adipose tissue
WD SW High-fat Western diet-fed
Wnt Wingless-related integration site

XO xanthine oxidase

ZEB1/2 Zinc finger E-box binding homeobox 1 and Zinc finger E-box binding homeobox 2

ZFP57 zinc finger protein 57

References

1. WHO. *Cancer*; World Health Organization: Geneva, Switzerland, 2024. Available online: https://www.who.int/news-room/fact-sheets/detail/cancer (accessed on 13 June 2025).

- 2. Hausman, D.M. What Is Cancer? Perspect. Biol. Med. 2019, 62, 778–784. [CrossRef] [PubMed]
- 3. Mumtaz, S.; Akhtar, N.; Ahmed, A.; Qazi, A.S. Dietary Pattern and Cancer. *Cancer Treat. Res.* **2024**, *191*, 191–216. [CrossRef] [PubMed]
- 4. Guasch-Ferré, M.; Willett, W.C. The Mediterranean diet and health: A comprehensive overview. *J. Intern. Med.* **2021**, 290, 549–566. [CrossRef] [PubMed]
- 5. Barber, T.M.; Kabisch, S.; Pfeiffer, A.F.H.; Weickert, M.O. The Effects of the Mediterranean Diet on Health and Gut Microbiota. *Nutrients* **2023**, *15*, 2150. [CrossRef] [PubMed]
- 6. Hanahan, D. Hallmarks of Cancer: New Dimensions. Cancer Discov. 2022, 12, 31–46. [CrossRef] [PubMed]
- 7. Levatić, J.; Salvadores, M.; Fuster-Tormo, F.; Supek, F. Mutational signatures are markers of drug sensitivity of cancer cells. *Nat. Commun.* **2022**, 25, 2926. [CrossRef] [PubMed]
- 8. Liu, Y.P.; Zheng, C.C.; Huang, Y.N.; He, M.L.; Xu, W.W.; Li, B. Molecular mechanisms of chemo- and radiotherapy resistance and the potential implications for cancer treatment. *MedComm* **2021**, *10*, 315–340. [CrossRef] [PubMed]
- 9. Cháirez-Ramírez, M.H.; de la Cruz-López, K.G.; García-Carrancá, A. Polyphenols as Antitumor Agents Targeting Key Players in Cancer-Driving Signaling Pathways. *Front. Pharmacol.* **2021**, 20, 710304. [CrossRef] [PubMed]
- 10. Luo, H.; Xia, X.; Huang, L.B.; An, H.; Cao, M.; Kim, G.D.; Chen, H.N.; Zhang, W.H.; Shu, Y.; Kong, X.; et al. Pan-cancer single-cell analysis reveals the heterogeneity and plasticity of cancer-associated fibroblasts in the tumor microenvironment. *Nat. Commun.* 2022, *4*, 6619. [CrossRef] [PubMed]
- 11. Musella, M.; Guarracino, A.; Manduca, N.; Galassi, C.; Ruggiero, E.; Potenza, A.; Maccafeo, E.; Manic, G.; Mattiello, L.; Soliman Abdel Rehim, S. Type I IFNs promote cancer cell stemness by triggering the epigenetic regulator KDM1B. *Nat. Immunol.* **2022**, 23, 1379–1392. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 31 of 41

12. Fanelli, G.N.; Naccarato, A.G.; Scatena, C. Recent Advances in Cancer Plasticity: Cellular Mechanisms, Surveillance Strategies, and Therapeutic Optimization. *Front. Oncol.* **2020**, 22, 569. [CrossRef] [PubMed]

- 13. Nyga, A.; Ganguli, S.; Matthews, H.K.; Baum, B. The role of RAS oncogenes in controlling epithelial mechanics. *Trends Cell Biol.* **2023**, *33*, 60–69. [CrossRef] [PubMed]
- 14. Lavoie, H.; Gagnon, J.; Therrien, M. ERK signalling: A master regulator of cell behaviour, life and fate. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 607–632. [CrossRef] [PubMed]
- 15. Bahar, M.E.; Kim, H.J.; Kim, D.R. Targeting the RAS/RAF/MAPK pathway for cancer therapy: From mechanism to clinical studies. *Signal Transduct. Target. Ther.* **2023**, *18*, 455. [CrossRef] [PubMed]
- 16. Cordani, M.; Dando, I.; Ambrosini, G.; González-Menéndez, P. Signaling, cancer cell plasticity, and intratumor heterogeneity. *Cell Commun. Signal.* **2024**, *3*, 255. [CrossRef] [PubMed]
- 17. Jolly, M.K.; Somarelli, J.A.; Sheth, M.; Biddle, A.; Tripathi, S.C.; Armstrong, A.J.; Hanash, S.M.; Bapat, S.A.; Rangarajan, A.; Levine, H. Hybrid epithelial/mesenchymal phenotypes promote metastasis and therapy resistance across carcinomas. *Pharmacol. Ther.* **2019**, *194*, 161–184. [CrossRef] [PubMed]
- 18. Kang, A.R.; Kim, J.L.; Kim, Y.; Kang, S.; Oh, S.C.; Park, J.K. A novel RIP1-mediated canonical WNT signaling pathway that promotes colorectal cancer metastasis via β -catenin stabilization-induced EMT. *Cancer Gene Ther.* **2023**, *30*, 1403–1413. [CrossRef] [PubMed]
- 19. Vasaikar, S.V.; Deshmukh, A.P.; den Hollander, P.; Addanki, S.; Kuburich, N.A.; Kudaravalli, S.; Joseph, R.; Chang, J.T.; Soundararajan, R.; Mani, S.A. EMTome: A resource for pan-cancer analysis of epithelial-mesenchymal transition genes and signatures. *Br. J. Cancer* 2021, 124, 259–269. [CrossRef] [PubMed]
- 20. Hoxhaj, G.; Manning, B.D. The PI3K-AKT network at the interface of oncogenic signalling and cancer metabolism. *Nat. Rev. Cancer* **2020**, 20, 74–88. [CrossRef] [PubMed]
- 21. Tan, Y.; Li, J.; Zhao, G.; Huang, K.C.; Cardenas, H.; Wang, Y.; Matei, D.; Cheng, J.X. Metabolic reprogramming from glycolysis to fatty acid uptake and beta-oxidation in platinum-resistant cancer cells. *Nat. Commun.* **2022**, *5*, 4554. [CrossRef] [PubMed]
- 22. Shah, D.D.; Chorawala, M.R.; Raghani, N.R.; Patel, R.; Fareed, M.; Kashid, V.A.; Prajapati, B.G. Tumor microenvironment: Recent advances in understanding and its role in modulating cancer therapies. *Med. Oncol.* **2025**, *42*, 117. [CrossRef] [PubMed]
- 23. Bejarano, L.; Jordāo, M.J.C.; Joyce, J.A. Therapeutic Targeting of the Tumor Microenvironment. *Cancer Discov.* **2021**, *11*, 933–959. [CrossRef] [PubMed]
- 24. Elinav, E.; Nowarski, R.; Thaiss, C.A.; Hu, B.; Jin, C.; Flavell, R.A. Inflammation-induced cancer: Crosstalk between tumours, immune cells and microorganisms. *Nat. Rev. Cancer* **2013**, *13*, 759–771. [CrossRef] [PubMed]
- 25. Karimi, M.; Asbaghi, O.; Hooshmand, F.; Aghayan, A.H.; Shariati, A.A.; Kazemi, K.; Amirpour, M.; Davoodi, S.H.; Larijani, B. Adherence to Mediterranean Diet and Breast Cancer Risk: A Meta-Analysis of Prospective Observational Studies. *Health Sci. Rep.* 2025, 8, 70736. [CrossRef] [PubMed]
- 26. Giordano, G.; Mastrantoni, L.; Terranova, R.; Colloca, G.F.; Zuccalà, G.; Landi, F. The role of Mediterranean diet in cancer incidence and mortality in the older adults. *NPJ Aging* **2024**, *10*, 61. [CrossRef] [PubMed]
- 27. Pan, B.; Wu, Y.; Yang, Q.; Ge, L.; Gao, C.; Xun, Y.; Tian, J.; Ding, G. The impact of major dietary patterns on glycemic control, cardiovascular risk factors, and weight loss in patients with type 2 diabetes: A network meta-analysis. *J. Evid. Based Med.* **2019**, 12, 29–39. [CrossRef] [PubMed]
- 28. Oppedisano, F.; Mollace, R.; Tavernese, A.; Gliozzi, M.; Musolino, V.; Macrì, R.; Carresi, C.; Maiuolo, J.; Serra, M.; Cardamone, A.; et al. PUFA Supplementation and Heart Failure: Effects on Fibrosis and Cardiac Remodeling. *Nutrients* **2021**, *13*, 2965. [CrossRef] [PubMed]
- 29. Tufarelli, V.; Casalino, E.; D'Alessandro, A.G.; Laudadio, V. Dietary Phenolic Compounds: Biochemistry, Metabolism and Significance in Animal and Human Health. *Curr. Drug Metab.* **2017**, *18*, 905–913. [CrossRef] [PubMed]
- 30. Rana, A.; Samtiya, M.; Dhewa, T.; Mishra, V.; Aluko, R.E. Health benefits of polyphenols: A concise review. *J. Food Biochem.* **2022**, 46, 14264. [CrossRef] [PubMed]
- 31. Williamson, G. Bioavailability of Food Polyphenols: Current State of Knowledge. *Annu. Rev. Food Sci. Technol.* **2025**, *16*, 315–332. [CrossRef] [PubMed]
- 32. Di Lorenzo, C.; Colombo, F.; Biella, S.; Stockley, C.; Restani, P. Polyphenols and Human Health: The Role of Bioavailability. Nutrients 2021, 13, 273. [CrossRef] [PubMed]
- 33. Bobrysheva, T.N.; Anisimov, G.S.; Zolotoreva, M.S.; Bobryshev, D.V.; Budkevich, R.O.; Moskalev, A.A. [Polyphenols as promising bioactive compounds]. *Vopr. Pitan.* **2023**, *92*, 92–107. [CrossRef] [PubMed]
- 34. Wan, S.; Luo, J.; Zhu, Y.; An, P.; Luo, Y.; Xing, Q. The Effect of Antioxidant Polyphenol Supplementation on Cardiometabolic Risk Factors: A Systematic Review and Meta-Analysis. *Nutrients* **2024**, *16*, 4206. [CrossRef] [PubMed]
- 35. Cao, H.; Ou, J.; Chen, L.; Zhang, Y.; Szkudelski, T.; Delmas, D.; Daglia, M.; Xiao, J. Dietary polyphenols and type 2 diabetes: Human Study and Clinical Trial. *Crit. Rev. Food Sci. Nutr.* **2019**, *59*, 3371–3379. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 32 of 41

36. Wang, S.; Du, Q.; Meng, X.; Zhang, Y. Natural polyphenols: A potential prevention and treatment strategy for metabolic syndrome. *Food Funct.* **2022**, *13*, 9734–9753. [CrossRef] [PubMed]

- 37. Jomova, K.; Alomar, S.Y.; Valko, R.; Liska, J.; Nepovimova, E.; Kuca, K.; Valko, M. Flavonoids and their role in oxidative stress, inflammation, and human diseases. *Chem. Biol. Interact.* **2025**, 25, 111489. [CrossRef] [PubMed]
- 38. González Mosquera, D.M.; Hernández Ortega, Y.; Fernández, P.L.; González, Y.; Doens, D.; Vander Heyden, Y.; Foubert, K.; Pieters, L. Flavonoids from Boldoa purpurascens inhibit proinflammatory cytokines (TNF-α and IL-6) and the expression of COX-2. *Phytother. Res.* **2018**, 32, 1750–1754. [CrossRef] [PubMed]
- 39. Singaravelan, N.; Tollefsbol, T.O. Polyphenol-Based Prevention and Treatment of Cancer Through Epigenetic and Combinatorial Mechanisms. *Nutrients* **2025**, *17*, 616. [CrossRef] [PubMed]
- 40. Benvenuto, M.; Albonici, L.; Focaccetti, C.; Ciuffa, S.; Fazi, S.; Cifaldi, L.; Miele, M.T.; De Maio, F.; Tresoldi, I.; Manzari, V.; et al. Polyphenol-Mediated Autophagy in Cancer: Evidence of In Vitro and in vivo Studies. *Int. J. Mol. Sci.* 2020, 21, 6635. [CrossRef] [PubMed]
- 41. Martemucci, G.; Khalil, M.; Di Luca, A.; Abdallah, H.; D'Alessandro, A.G. Comprehensive Strategies for Metabolic Syndrome: How Nutrition, Dietary Polyphenols, Physical Activity, and Lifestyle Modifications Address Diabesity, Cardiovascular Diseases, and Neurodegenerative Conditions. *Metabolites* 2024, 14, 327. [CrossRef] [PubMed]
- 42. Roszkowska, M. Multilevel Mechanisms of Cancer Drug Resistance. Int. J. Mol. Sci. 2024, 25, 12402. [CrossRef] [PubMed]
- 43. Auti, A.; Tathode, M.; Marino, M.M.; Vitiello, A.; Ballini, A.; Miele, F.; Mazzone, V.; Ambrosino, A.; Boccellino, M. Nature's weapons: Bioactive compounds as anti-cancer agents. *AIMS Public Health* **2024**, *11*, 747–772. [CrossRef] [PubMed]
- 44. Pierdomenico, M.; Cicero, A.F.G.; Veronesi, M.; Fogacci, F.; Riccioni, C.; Benassi, B. Effect of Citrus bergamia extract on lipid profile: A combined in vitro and human study. *Phytother. Res.* **2023**, *37*, 4185–4195. [CrossRef] [PubMed]
- 45. Cordeiro, K.C.; Scaffo, J.; Flexa, B.N.; Gama, C.C.A.; Ferreira, M.A.; Cruz, R.A.S.; Aguiar-Alves, F.; Rocha, L.; Machado, F.P.; Fernandes, C.P. Characterization of bergamot essential oil: Chemical, microbiological and colloidal aspects. *Braz. J. Biol.* **2024**, *26*, 275622. [CrossRef] [PubMed]
- 46. Mollace, V.; Scicchitano, M.; Paone, S.; Casale, F.; Calandruccio, C.; Gliozzi, M.; Musolino, V.; Carresi, C.; Maiuolo, J.; Nucera, S.; et al. Hypoglycemic and Hypolipemic Effects of a New Lecithin Formulation of Bergamot Polyphenolic Fraction: A Double Blind, Randomized, Placebo- Controlled Study. Endocr. Metab. Immune Disord.-Drug Targets 2019, 19, 136–143. [CrossRef] [PubMed]
- 47. Mollace, V.; Sacco, I.; Janda, E.; Malara, C.; Ventrice, D.; Colica, C.; Visalli, V.; Muscoli, S.; Ragusa, S.; Muscoli, C.; et al. Hypolipemic and hypoglycaemic activity of bergamot polyphenols: From animal models to human studies. *Fitoterapia* **2011**, *82*, 309–316. [CrossRef] [PubMed]
- 48. Bavaro, A.R.; Tarantini, A.; Bruno, A.; Logrieco, A.F.; Gallo, A.; Mita, G.; Valerio, F.; Bleve, G.; Cardinali, A. Functional foods in Mediterranean diet: Exploring the functional features of vegetable case-studies obtained also by biotechnological approaches. *Aging Clin. Exp. Res.* 2024, 36, 208. [CrossRef] [PubMed]
- 49. Ding, X.; Chen, Y.; Du, L.; Li, J.; Meng, X.; Lv, H.; Tong, B.; Niu, G.; Jian, T.; Chen, J. Benefits of inulin and fructo-oligosaccharides on high fat diet-induced type 2 diabetes mellitus by regulating the gut microbiota in mice. *J. Nutr. Biochem.* **2025**, 141, 109908. [CrossRef] [PubMed]
- 50. Tsimihodimos, V.; Psoma, O. Extra Virgin Olive Oil and Metabolic Diseases. Int. J. Mol. Sci. 2024, 25, 8117. [CrossRef] [PubMed]
- 51. Maruca, A.; Catalano, R.; Bagetta, D.; Mesiti, F.; Ambrosio, F.A.; Romeo, I.; Moraca, F.; Rocca, R.; Ortuso, F.; Artese, A.; et al. The Mediterranean Diet as source of bioactive compounds with multi-targeting anti-cancer profile. *Eur. J. Med. Chem.* **2019**, *181*, 111579. [CrossRef] [PubMed]
- 52. Lammi, C.; Bellumori, M.; Cecchi, L.; Bartolomei, M.; Bollati, C.; Clodoveo, M.L.; Corbo, F.; Arnoldi, A.; Mulinacci, N. Extra Virgin Olive Oil Phenol Extracts Exert Hypocholesterolemic Effects through the Modulation of the LDLR Pathway: In Vitro and Cellular Mechanism of Action Elucidation. *Nutrients* 2020, 12, 1723. [CrossRef] [PubMed]
- 53. Rishmawi, S.; Haddad, F.; Dokmak, G.; Karaman, R. A Comprehensive Review on the Anti-Cancer Effects of Oleuropein. *Life* **2022**, *12*, 1140. [CrossRef] [PubMed]
- 54. Alizadeh, S.R.; Ebrahimzadeh, M.A. Quercetin derivatives: Drug design, development, and biological activities, a review. *Eur. J. Med. Chem.* **2022**, 229, 114068. [CrossRef] [PubMed]
- 55. Ileriturk, M.; Kandemir, O.; Kandemir, F.M. Evaluation of protective effects of quercetin against cypermethrin-induced lung toxicity in rats via oxidative stress, inflammation, apoptosis, autophagy, and endoplasmic reticulum stress pathway. *Environ. Toxicol.* 2022, 37, 2639–2650. [CrossRef] [PubMed]
- Wu, S.X.; Xiong, R.G.; Huang, S.Y.; Zhou, D.D.; Saimaiti, A.; Zhao, C.N.; Shang, A.; Zhang, Y.J.; Gan, R.Y.; Li, H.B. Effects and mechanisms of resveratrol for prevention and management of cancers: An updated review. Crit. Rev. Food Sci. Nutr. 2023, 63, 12422–12440. [CrossRef] [PubMed]
- 57. Nani, A.; Murtaza, B.; Sayed Khan, A.; Khan, N.A.; Hichami, A. Antioxidant and Anti-Inflammatory Potential of Polyphenols Contained in Mediterranean Diet in Obesity: Molecular Mechanisms. *Molecules* **2021**, *26*, 985. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 33 of 41

58. Cheng, H.; Zhang, D.; Wu, J.; Liu, J.; Zhou, Y.; Tan, Y.; Feng, W.; Peng, C. Interactions between gut microbiota and polyphenols: A mechanistic and metabolomic review. *Phytomedicine* **2023**, *119*, 154979. [CrossRef] [PubMed]

- 59. Ed-Dahmani, I.; El Fadili, M.; Nouioura, G.; Kandsi, F.; Atki, Y.E.; Abuelizz, H.A.; Conte, R.; Zahra Lafdil, F.; Taleb, A.; Abdellaoui, A.; et al. *Ferula communis* leaf extract: Antioxidant capacity, UHPLC-MS/MS analysis, and in vivo and In Silico toxicity investigations. *Front. Chem.* **2025**, *12*, 1485463. [CrossRef] [PubMed]
- 60. Macrì, R.; Maiuolo, J.; Scarano, F.; Musolino, V.; Fregola, A.; Gliozzi, M.; Carresi, C.; Nucera, S.; Serra, M.; Caminiti, R.; et al. Evaluation of the Potential Beneficial Effects of *Ferula communis* L. Extract Supplementation in Postmenopausal Discomfort. *Nutrients* 2024, 16, 2651. [CrossRef] [PubMed]
- 61. Salerno, R.; Casale, F.; Calandruccio, C.; Procopio, A. Characterization of flavonoids in Citrus bergamia (Bergamot) polyphenolic fraction by liquid chromatography–high resolution mass spectrometry (LC/HRMS). *PharmaNutrition* **2016**, *4*, 2213–4344. [CrossRef]
- 62. Buzzanca, C.; Di Stefano, V.; D'Amico, A.; Gallina, A.; Melilli, M.G. A systematic review on *Cynara cardunculus* L.: Bioactive compounds, nutritional properties and food-industry applications of a sustainable food. *Nat. Prod. Res.* **2024**, *3*, 20. [CrossRef] [PubMed]
- 63. Li, X.; Muñoz-Díez, C.; Miho, H.; Zhang, L.; Li, P.; Priego, F.; Oulbi, S.; Uyanik, E.B.; Koubouris, G.; Perri, E.; et al. Evaluation of phenolics in the analysis of virgin olive oil using near infrared spectroscopy. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* 2025, 326, 125262. [CrossRef] [PubMed]
- 64. Ussia, S.; Ritorto, G.; Mollace, R.; Serra, M.; Tavernese, A.; Altomare, C.; Muscoli, C.; Fini, M.; Barillà, F.; Indolfi, C.; et al. Exploring the Benefits of Extra Virgin Olive Oil on Cardiovascular Health Enhancement and Disease Prevention: A Systematic Review. *Nutrients* 2025, 17, 1843. [CrossRef] [PubMed]
- 65. Sun, W.; Shahrajabian, M.H. Therapeutic Potential of Phenolic Compounds in Medicinal Plants-Natural Health Products for Human Health. *Molecules* **2023**, *28*, 1845. [CrossRef] [PubMed]
- 66. Leischner, C.; Burkard, M.; Michel, A.; Berchtold, S.; Niessner, H.; Marongiu, L.; Busch, C.; Frank, J.; Lauer, U.M.; Venturelli, S. Comparative Analysis of the Antitumor Activity of Cis- and Trans-Resveratrol in Human Cancer Cells with Different p53 Status. *Molecules* 2021, 26, 5586. [CrossRef] [PubMed]
- 67. Tian, B.; Liu, J. Resveratrol: A review of plant sources, synthesis, stability, modification and food application. *J. Sci. Food Agric.* **2020**, *100*, 1392–1404. [CrossRef] [PubMed]
- 68. Nouioura, G.; El Fadili, M.; El Barnossi, A.; Loukili, E.H.; Laaroussi, H.; Bouhrim, M.; Giesy, J.P.; Aboul-Soud, M.A.M.; Al-Sheikh, Y.A.; Lyoussi, B.; et al. Comprehensive analysis of different solvent extracts of Ferula communis L. fruit reveals phenolic compounds and their biological properties via in vitro and in silico assays. *Sci. Rep.* **2024**, *14*, 8325. [CrossRef] [PubMed]
- 69. Sonigra, P.; Meena, M. Metabolic Profile, Bioactivities, and Variations in the Chemical Constituents of Essential Oils of the *Ferula Genus* (Apiaceae). *Front. Pharmacol.* **2021**, *11*, 608649. [CrossRef] [PubMed]
- 70. Louvet, M.S.; Gault, G.; Lefebvre, S.; Popowycz, F.; Boulven, M.; Besse, S.; Benoit, E.; Lattard, V.; Grancher, D. Comparative inhibitory effect of prenylated coumarins, ferulenol and ferprenin, contained in the 'poisonous chemotype' of Ferula communis on mammal liver microsomal VKORC1 activity. *Phytochemistry* **2015**, *118*, 124–130. [CrossRef] [PubMed]
- 71. Naji Reyhani Garmroudi, S.; Karimi, E.; Oskoueian, E.; Homayouni-Tabrizi, M.; Iranshahi, M. Ferutinin: A phytoestrogen from ferula and its anticancer, antioxidant, and toxicity properties. *J. Biochem. Mol. Toxicol.* **2021**, *35*, 22713. [CrossRef] [PubMed]
- 72. Nolfi-Donegan, D.; Braganza, A.; Shiva, S. Mitochondrial electron transport chain: Oxidative phosphorylation, oxidant production, and methods of measurement. *Redox Biol.* **2020**, *37*, 101674. [CrossRef] [PubMed]
- 73. Jelic, M.D.; Mandic, A.D.; Maricic, S.M.; Srdjenovic, B.U. Oxidative stress and its role in cancer. *J. Cancer Res. Ther.* **2021**, 17, 22–28. [CrossRef] [PubMed]
- 74. Wu, Q.; Yao, J.; Xiao, M.; Zhang, X.; Zhang, M.; Xi, X. Targeting Nrf2 signaling pathway: New therapeutic strategy for cardiovascular diseases. *J. Drug Target*. **2024**, 32, 874–883. [CrossRef] [PubMed]
- 75. Sciaccotta, R.; Gangemi, S.; Penna, G.; Giordano, L.; Pioggia, G.; Allegra, A. Potential New Therapies "ROS-Based" in CLL: An Innovative Paradigm in the Induction of Tumor Cell Apoptosis. *Antioxidants* **2024**, *13*, 475. [CrossRef] [PubMed]
- 76. Altomare, A.; Baron, G.; Gianazza, E.; Banfi, C.; Carini, M.; Aldini, G. Lipid peroxidation derived reactive carbonyl species in free and conjugated forms as an index of lipid peroxidation: Limits and perspectives. *Redox Biol.* **2021**, *42*, 101899. [CrossRef] [PubMed]
- 77. Jomova, K.; Raptova, R.; Alomar, S.Y.; Alwasel, S.H.; Nepovimova, E.; Kuca, K.; Valko, M. Reactive oxygen species, toxicity, oxidative stress, and antioxidants: Chronic diseases and aging. *Arch. Toxicol.* **2023**, *97*, 2499–2574. [CrossRef] [PubMed]
- 78. Guo, H.; Yang, Y.; Lou, Y.; Zuo, Z.; Cui, H.; Deng, H.; Zhu, Y.; Fang, J. Apoptosis and DNA damage mediated by ROS involved in male reproductive toxicity in mice induced by Nickel. *Ecotoxicol. Environ. Saf.* **2023**, 268, 115679. [CrossRef] [PubMed]
- Chen, B.; Lu, M.; Chen, Q.; Zou, E.; Bo, Z.; Li, J.; Zhao, R.; Zhao, J.; Yu, Z.; Chen, G.; et al. Systematic profiling of mitochondriarelated transcriptome in tumorigenesis, prognosis, and tumor immune microenvironment of intrahepatic cholangiocarcinoma: A multi-center cohort study. Front. Genet. 2024, 15, 1430885. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 34 of 41

80. Ávila-Escalante, M.L.; Coop-Gamas, F.; Cervantes-Rodríguez, M.; Méndez-Iturbide, D.; Aranda-González, I.I. The effect of diet on oxidative stress and metabolic diseases-Clinically controlled trials. *J. Food Biochem.* **2020**, 44, 13191. [CrossRef] [PubMed]

- 81. Kong, C.Y.; Guo, Z.; Song, P.; Zhang, X.; Yuan, Y.P.; Teng, T.; Yan, L.; Tang, Q.Z. Underlying the Mechanisms of Doxorubicin-Induced Acute Cardiotoxicity: Oxidative Stress and Cell Death. *Int. J. Biol. Sci.* **2022**, *18*, 760–770. [CrossRef] [PubMed]
- 82. de Almeida, A.J.P.O.; de Oliveira, J.C.P.L.; da Silva Pontes, L.V.; de Souza Júnior, J.F.; Gonçalves, T.A.F.; Dantas, S.H.; de Almeida Feitosa, M.S.; Silva, A.O.; de Medeiros, I.A. ROS: Basic Concepts, Sources, Cellular Signaling, and its Implications in Aging Pathways. *Oxid. Med. Cell Longev.* 2022, 19, 1225578. [CrossRef] [PubMed]
- 83. Vašková, J.; Kočan, L.; Vaško, L.; Perjési, P. Glutathione-Related Enzymes and Proteins: A Review. *Molecules* **2023**, *28*, 1447. [CrossRef] [PubMed]
- 84. Bianchi, E.; Erbasan, H.; Riso, P.; Perna, S. Impact of the Mediterranean Diet on Athletic Performance, Muscle Strength, Body Composition, and Antioxidant Markers in Both Athletes and Non-Professional Athletes: A Systematic Review of Intervention Trials. *Nutrients* 2024, 16, 3454. [CrossRef] [PubMed]
- 85. Zhu, R.; Fogelholm, M.; Poppitt, S.D.; Silvestre, M.P.; Møller, G.; Huttunen-Lenz, M.; Stratton, G.; Sundvall, J.; Råman, L.; Jalo, E.; et al. Adherence to a Plant-Based Diet and Consumption of Specific Plant Foods-Associations with 3-Year Weight-Loss Maintenance and Cardiometabolic Risk Factors: A Secondary Analysis of the PREVIEW Intervention Study. *Nutrients* **2021**, *13*, 3916. [CrossRef] [PubMed]
- 86. La Scala, S.; Naselli, F.; Quatrini, P.; Gallo, G.; Caradonna, F. Drought-Adapted Mediterranean Diet Plants: A Source of Bioactive Molecules Able to Give Nutrigenomic Effects per sè or to Obtain Functional Foods. *Int. J. Mol. Sci.* **2024**, 25, 2235. [CrossRef] [PubMed]
- 87. Nazari, A.; Osati, P.; Seifollahy Fakhr, S.; Faghihkhorasani, F.; Ghanaatian, M.; Faghihkhorasani, F.; Rezaei-Tazangi, F.; Pazhouhesh Far, N.; Shourideh, A.; Ebrahimi, N.; et al. New Emerging Therapeutic Strategies Based on Manipulation of the Redox Regulation Against Therapy Resistance in Cancer. *Antioxid. Redox Signal.* **2024**. [CrossRef] [PubMed]
- 88. Yuan, M.; Zhang, G.; Bai, W.; Han, X.; Li, C.; Bian, S. The Role of Bioactive Compounds in Natural Products Extracted from Plants in Cancer Treatment and Their Mechanisms Related to Anticancer Effects. *Oxid. Med. Cell Longev.* **2022**, *15*, 1429869. [CrossRef] [PubMed]
- 89. Trinh, V.H.; Nguyen Huu, T.; Sah, D.K.; Choi, J.M.; Yoon, H.J.; Park, S.C.; Jung, Y.S.; Lee, S.R. Redox Regulation of PTEN by Reactive Oxygen Species: Its Role in Physiological Processes. *Antioxidants* **2024**, *13*, 199. [CrossRef] [PubMed]
- 90. Ghofrani-Shahpar, M.; Pakravan, K.; Razmara, E.; Amooie, F.; Mahmoudian, M.; Heshmati, M.; Babashah, S. Cancer-associated fibroblasts drive colorectal cancer cell progression through exosomal miR-20a-5p-mediated targeting of PTEN and stimulating interleukin-6 production. *BMC Cancer* **2024**, 24, 400. [CrossRef] [PubMed]
- 91. Glaviano, A.; Foo, A.S.C.; Lam, H.Y.; Yap, K.C.H.; Jacot, W.; Jones, R.H.; Eng, H.; Nair, M.G.; Makvandi, P.; Geoerger, B.; et al. PI3K/AKT/mTOR signaling transduction pathway and targeted therapies in cancer. *Mol. Cancer.* **2023**, 22, 138. [CrossRef] [PubMed]
- 92. Mohideen, K.; Chandrasekaran, K.; Kareema, M.; Jeyanthikumari, T.; Dhungel, S.; Ghosh, S. Assessment of Antioxidant Enzyme Superoxide Dismutase (SOD) in Oral Cancer: Systematic Review and Meta-Analysis. *Dis. Markers* **2024**, *16*, 2264251. [CrossRef] [PubMed]
- 93. Liu, Y.; Su, Z.; Tavana, O.; Gu, W. Understanding the complexity of p53 in a new era of tumor suppression. *Cancer Cell.* **2024**, 42, 946–967. [CrossRef] [PubMed]
- 94. Suzuki, T.; Takahashi, J.; Yamamoto, M. Molecular Basis of the KEAP1-NRF2 Signaling Pathway. *Mol. Cells.* **2023**, *46*, 133–141. [CrossRef] [PubMed]
- 95. Tülüce, Y.; Keleş, A.Y.; Köstekci, S. Assessment of redox homeostasis via genotoxicity, cytotoxicity, apoptosis and NRF-2 in colorectal cancer cell lines after treatment with *Ganoderma lucidum* extract. *Drug Chem. Toxicol.* **2024**, 47, 693–709. [CrossRef] [PubMed]
- 96. Averill-Bates, D. Reactive oxygen species and cell signaling. Review. *Biochim. Biophys. Acta Mol. Cell Res.* **2024**, 1871, 119573. [CrossRef] [PubMed]
- 97. Adinolfi, S.; Patinen, T.; Jawahar Deen, A.; Pitkänen, S.; Härkönen, J.; Kansanen, E.; Küblbeck, J.; Levonen, A.L. The KEAP1-NRF2 pathway: Targets for therapy and role in cancer. *Redox Biol.* **2023**, *63*, 102726. [CrossRef] [PubMed]
- 98. Moon, H.; Ro, S.W. MAPK/ERK Signaling Pathway in Hepatocellular Carcinoma. Cancers 2021, 13, 3026. [CrossRef] [PubMed]
- 99. Chen, J.Y.; Yang, Y.J.; Meng, X.Y.; Lin, R.H.; Tian, X.Y.; Zhang, Y.; Lai, W.F.; Yang, C.; Ma, X.Q.; Huang, M.Q. Oxysophoridine inhibits oxidative stress and inflammation in hepatic fibrosis via regulating Nrf2 and NF-κB pathways. *Phytomedicine* **2024**, 132, 155585. [CrossRef] [PubMed]
- 100. Zhou, X.; An, B.; Lin, Y.; Ni, Y.; Zhao, X.; Liang, X. Molecular mechanisms of ROS-modulated cancer chemoresistance and therapeutic strategies. *Biomed. Pharmacother.* **2023**, *165*, 115036. [CrossRef] [PubMed]
- 101. Fleming Martinez, A.K.; Storz, P. Protein kinase D1—A targetable mediator of pancreatic cancer development. *Biochim. Biophys. Acta Mol. Cell Res.* **2024**, *1871*, 119646. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 35 of 41

102. Li, Y.; Zhao, B.; Peng, J.; Tang, H.; Wang, S.; Peng, S.; Ye, F.; Wang, J.; Ouyang, K.; Li, J.; et al. Inhibition of NF-κB signaling unveils novel strategies to overcome drug resistance in cancers. *Drug Resist. Updat.* **2024**, *73*, 101042. [CrossRef] [PubMed]

- 103. Iqbal, M.J.; Kabeer, A.; Abbas, Z.; Siddiqui, H.A.; Calina, D.; Sharifi-Rad, J.; Cho, W.C. Interplay of oxidative stress, cellular communication and signaling pathways in cancer. *Cell Commun. Signal.* **2024**, 22, 7. [CrossRef] [PubMed]
- 104. De Rosa, M.; Johnson, S.A.; Opresko, P.L. Roles for the 8-Oxoguanine DNA Repair System in Protecting Telomeres From Oxidative Stress. *Front. Cell Dev. Biol.* **2021**, *9*, 758402. [CrossRef] [PubMed]
- 105. Verma, P.; Rishi, B.; George, N.G.; Kushwaha, N.; Dhandha, H.; Kaur, M.; Jain, A.; Jain, A.; Chaudhry, S.; Singh, A.; et al. Recent advances and future directions in etiopathogenesis and mechanisms of reactive oxygen species in cancer treatment. *Pathol. Oncol. Res.* 2023, 29, 1611415. [CrossRef] [PubMed]
- 106. Morse, P.T.; Arroum, T.; Wan, J.; Pham, L.; Vaishnav, A.; Bell, J.; Pavelich, L.; Malek, M.H.; Sanderson, T.H.; Edwards, B.F.P.; et al. Phosphorylations and Acetylations of Cytochrome *c* Control Mitochondrial Respiration, Mitochondrial Membrane Potential, Energy, ROS, and Apoptosis. *Cells* **2024**, *13*, 493. [CrossRef] [PubMed]
- 107. Mustafa, M.; Ahmad, R.; Tantry, I.Q.; Ahmad, W.; Siddiqui, S.; Alam, M.; Abbas, K.; Moinuddin; Hassan, M.I.; Habib, S.; et al. Apoptosis: A Comprehensive Overview of Signaling Pathways, Morphological Changes, and Physiological Significance and Therapeutic Implications. *Cells* 2024, 13, 1838. [CrossRef] [PubMed]
- 108. Algieri, C.; Bernardini, C.; Oppedisano, F.; La Mantia, D.; Trombetti, F.; Palma, E.; Forni, M.; Mollace, V.; Romeo, G.; Nesci, S. Mitochondria Bioenergetic Functions and Cell Metabolism Are Modulated by the Bergamot Polyphenolic Fraction. *Cells* **2022**, *11*, 1401. [CrossRef] [PubMed]
- 109. Mollace, R.; Macrì, R.; Tavernese, A.; Gliozzi, M.; Musolino, V.; Carresi, C.; Maiuolo, J.; Fini, M.; Volterrani, M.; Mollace, V. Comparative Effect of Bergamot Polyphenolic Fraction and Red Yeast Rice Extract in Rats Fed a Hyperlipidemic Diet: Role of Antioxidant Properties and PCSK9 Expression. *Nutrients* 2022, 14, 477. [CrossRef] [PubMed]
- 110. Mollace, R.; Macrì, R.; Nicita, M.; Musolino, V.; Gliozzi, M.; Carresi, C.; Bava, I.; Maiuolo, J.; Tavernese, A.; Cardamone, A.; et al. Bergamot Polyphenolic Extract Combined with Albedo and Pulp Fibres Counteracts Changes in Gut Microbiota Associated with High-Fat Diet: Implications for Lipoprotein Size Re-Arrangement. *Int. J. Mol. Sci.* 2023, 24, 12967. [CrossRef] [PubMed]
- 111. Gliozzi, M.; Walker, R.; Muscoli, S.; Vitale, C.; Gratteri, S.; Carresi, C.; Musolino, V.; Russo, V.; Janda, E.; Ragusa, S.; et al. Bergamot polyphenolic fraction enhances rosuvastatin-induced effect on LDL-cholesterol, LOX-1 expression and protein kinase B phosphorylation in patients with hyperlipidemia. *Int. J. Cardiol.* 2013, 170, 140–145. [CrossRef] [PubMed]
- 112. Musolino, V.; Gliozzi, M.; Scarano, F.; Bosco, F.; Scicchitano, M.; Nucera, S.; Carresi, C.; Ruga, S.; Zito, M.C.; Maiuolo, J.; et al. Bergamot Polyphenols Improve Dyslipidemia and Pathophysiological Features in a Mouse Model of Non-Alcoholic Fatty Liver Disease. *Sci. Rep.* 2020, *10*, 2565. [CrossRef] [PubMed]
- 113. Carresi, C.; Musolino, V.; Gliozzi, M.; Maiuolo, J.; Mollace, R.; Nucera, S.; Maretta, A.; Sergi, D.; Muscoli, S.; Gratteri, S.; et al. Anti-oxidant effect of bergamot polyphenolic fraction counteracts doxorubicin-induced cardiomyopathy: Role of autophagy and c-kit^{pos}CD45^{neg}CD31^{neg} cardiac stem cell activation. *J. Mol. Cell Cardiol.* 2018, 119, 10–18. [CrossRef] [PubMed]
- 114. Masala, V.; Jokić, S.; Aladić, K.; Molnar, M.; Casula, M.; Tuberoso, C.I.G. Chemical Profiling and Evaluation of Antioxidant Activity of Artichoke (*Cynara cardunculus* var. *scolymus*) Leaf By-Products' Extracts Obtained with Green Extraction Techniques. *Molecules* 2024, 29, 4816. [CrossRef]
- 115. Acquaviva, R.; Malfa, G.A.; Santangelo, R.; Bianchi, S.; Pappalardo, F.; Taviano, M.F.; Miceli, N.; Di Giacomo, C.; Tomasello, B. Wild Artichoke (*Cynara cardunculus* subsp. *sylvestris*, Asteraceae) Leaf Extract: Phenolic Profile and Oxidative Stress Inhibitory Effects on HepG2 Cells. *Molecules* 2023, 28, 2475. [CrossRef]
- 116. D'Antuono, I.; Garbetta, A.; Linsalata, V.; Minervini, F.; Cardinali, A. Polyphenols from artichoke heads (Cynara cardunculus (L.) subsp. scolymus Hayek): In vitro bio-accessibility, intestinal uptake and bioavailability. *Food Funct.* **2015**, *6*, 1268-77. [CrossRef] [PubMed]
- 117. Cho, J.Y.; Kim, A.R.; Jung, J.H.; Chun, T.; Rhee, M.H.; Yoo, E.S. Cytotoxic and pro-apoptotic activities of cynaropicrin, a sesquiterpene lactone, on the viability of leukocyte cancer cell lines. *Eur. J. Pharmacol.* **2004**, 492, 85–94. [CrossRef] [PubMed]
- 118. Liu, T.; Zhang, J.; Han, X.; Xu, J.; Wu, Y.; Fang, J. Promotion of HeLa cells apoptosis by cynaropicrin involving inhibition of thioredoxin reductase and induction of oxidative stress. *Free Radic. Biol. Med.* **2019**, *135*, 216–226. [CrossRef] [PubMed]
- 119. De Cicco, P.; Busà, R.; Ercolano, G.; Formisano, C.; Allegra, M.; Taglialatela-Scafati, O.; Ianaro, A. Inhibitory effects of cynaropicrin on human melanoma progression by targeting MAPK, NF-κB, and Nrf-2 signaling pathways in vitro. *Phytother. Res.* **2021**, *35*, 1432–1442. [CrossRef] [PubMed]
- 120. Jin, T.; Leng, B. Cynaropicrin Averts the Oxidative Stress and Neuroinflammation in Ischemic/Reperfusion Injury Through the Modulation of NF-kB. *Appl. Biochem. Biotechnol.* **2023**, *195*, 5424–5438. [CrossRef] [PubMed]
- 121. Nucera, S.; Scarano, F.; Macrì, R.; Mollace, R.; Gliozzi, M.; Carresi, C.; Ruga, S.; Serra, M.; Tavernese, A.; Caminiti, R.; et al. The Effect of an Innovative Combination of Bergamot Polyphenolic Fraction and *Cynara cardunculus* L. Extract on Weight Gain Reduction and Fat Browning in Obese Mice. *Int. J. Mol. Sci.* 2024, 25, 191. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 36 of 41

122. Musolino, V.; Gliozzi, M.; Bombardelli, E.; Nucera, S.; Carresi, C.; Maiuolo, J.; Mollace, R.; Paone, S.; Bosco, F.; Scarano, F.; et al. The synergistic effect of *Citrus bergamia* and *Cynara cardunculus* extracts on vascular inflammation and oxidative stress in non-alcoholic fatty liver disease. *J. Tradit. Complement. Med.* 2020, 10, 268–274. [CrossRef] [PubMed]

- 123. Bulotta, S.; Corradino, R.; Celano, M.; D'Agostino, M.; Maiuolo, J.; Oliverio, M.; Procopio, A.; Iannone, M.; Rotiroti, D.; Russo, D. Antiproliferative and antioxidant effects on breast cancer cells of oleuropein and its semisynthetic peracetylated derivatives. *Food Chem.* 2011, 127, 0308–8146. [CrossRef]
- 124. Liman, R.; Çoban, F.K.; Ciğerci, I.H.; Ibrahim, B.; Bozkurt, S. Antiangiogenic and Apoptotic Effects of Oleuropein on Breast Cancer Cells. *J. Pharm. Res.* 2017, 16, 1–10. [CrossRef]
- 125. Yan, C.M.; Chai, E.Q.; Cai, H.Y.; Miao, G.Y.; Ma, W. Oleuropein induces apoptosis via activation of caspases and suppression of phosphatidylinositol 3-kinase/protein kinase B pathway in HepG2 human hepatoma cell line. *Mol. Med. Rep.* 2015, 11, 4617–4624. [CrossRef] [PubMed]
- 126. Scicchitano, S.; Vecchio, E.; Battaglia, A.M.; Oliverio, M.; Nardi, M.; Procopio, A.; Costanzo, F.; Biamonte, F.; Faniello, M.C. The Double-Edged Sword of Oleuropein in Ovarian Cancer Cells: From Antioxidant Functions to Cytotoxic Effects. *Int. J. Mol. Sci.* 2023, 24, 842. [CrossRef]
- 127. Maiuolo, J.; Bava, I.; Carresi, C.; Gliozzi, M.; Musolino, V.; Scarano, F.; Nucera, S.; Scicchitano, M.; Bosco, F.; Ruga, S.; et al. The Effects of Bergamot Polyphenolic Fraction, *Cynara cardunculus* and *Olea europea* L. Extract on Doxorubicin-Induced Cardiotoxicity. *Nutrients* 2021, *13*, 2158. [CrossRef] [PubMed]
- 128. Algieri, C.; Bernardini, C.; Oppedisano, F.; La Mantia, D.; Trombetti, F.; Palma, E.; Forni, M.; Mollace, V.; Romeo, G.; Troisio, I.; et al. The Impairment of Cell Metabolism by Cardiovascular Toxicity of Doxorubicin Is Reversed by Bergamot Polyphenolic Fraction Treatment in Endothelial Cells. *Int. J. Mol. Sci.* 2022, 23, 8977. [CrossRef] [PubMed]
- 129. Maiuolo, J.; Carresi, C.; Gliozzi, M.; Musolino, V.; Scarano, F.; Coppoletta, A.R.; Guarnieri, L.; Nucera, S.; Scicchitano, M.; Bosco, F.; et al. Effects of Bergamot Polyphenols on Mitochondrial Dysfunction and Sarcoplasmic Reticulum Stress in Diabetic Cardiomyopathy. *Nutrients* 2021, *13*, 2476. [CrossRef] [PubMed]
- 130. Tuerxun, H.; Zhao, Y.; Li, Y.; Liu, X.; Wen, S.; Zhao, Y. Resveratrol alleviates testicular toxicity induced by anti-PD-1 through regulating the NRF2-SLC7A11-GPX4 pathway. *Front. Immunol.* **2025**, *16*, 1529991. [CrossRef] [PubMed]
- 131. Biswas, P.; Dey, D.; Biswas, P.K.; Rahaman, T.I.; Saha, S.; Parvez, A.; Khan, D.A.; Lily, N.J.; Saha, K.; Sohel, M.; et al. A Comprehensive Analysis and Anti-Cancer Activities of Quercetin in ROS-Mediated Cancer and Cancer Stem Cells. *Int. J. Mol. Sci.* 2022, 23, 11746. [CrossRef] [PubMed]
- 132. Dewanjee, S.; Bhattacharya, H.; Bhattacharyya, C.; Chakraborty, P.; Fleishman, J.; Alexiou, A.; Papadakis, M.; Jha, S.K. Nrf2/Keap1/ARE regulation by plant secondary metabolites: A new horizon in brain tumor management. *Cell Commun. Signal.* 2024, 15, 497. [CrossRef] [PubMed]
- 133. Guan, G.; Chen, Y.; Dong, Y. Unraveling the AMPK-SIRT1-FOXO Pathway: The In-Depth Analysis and Breakthrough Prospects of Oxidative Stress-Induced Diseases. *Antioxidants* **2025**, *14*, 70. [CrossRef] [PubMed]
- 134. Rajesh, R.U.; Sangeetha, D. Therapeutic potentials and targeting strategies of quercetin on cancer cells: Challenges and future prospects. *Phytomedicine* **2024**, 133, 155902. [CrossRef] [PubMed]
- 135. Khattar, S.; Khan, S.A.; Zaidi, S.A.A.; Darvishikolour, M.; Farooq, U.; Naseef, P.P.; Kurunian, M.S.; Khan, M.Z.; Shamim, A.; Khan, M.M.U.; et al. Resveratrol from Dietary Supplement to a Drug Candidate: An Assessment of Potential. *Pharmaceuticals* **2022**, 15, 957. [CrossRef] [PubMed]
- 136. Sitarek, P.; Merecz-Sadowska, A.; Sikora, J.; Dudzic, M.; Wiertek-Płoszaj, N.; Picot, L.; Śliwiński, T.; Kowalczyk, T. Flavonoids and their derivatives as DNA topoisomerase inhibitors with anti-cancer activity in various cell models: Exploring a novel mode of action. *Pharmacol. Res.* **2024**, 209, 107457. [CrossRef] [PubMed]
- 137. Macrì, R.; Musolino, V.; Gliozzi, M.; Carresi, C.; Maiuolo, J.; Nucera, S.; Scicchitano, M.; Bosco, F.; Scarano, F.; Ruga, S.; et al. *Ferula* L. Plant Extracts and Dose-Dependent Activity of Natural Sesquiterpene Ferutinin: From Antioxidant Potential to Cytotoxic Effects. *Molecules* 2020, 25, 5768. [CrossRef] [PubMed]
- 138. Macrì, R.; Bava, I.; Scarano, F.; Mollace, R.; Musolino, V.; Gliozzi, M.; Greco, M.; Foti, D.; Tucci, L.; Maiuolo, J.; et al. In Vitro Evaluation of Ferutinin Rich-Ferula communis L., ssp. glauca, Root Extract on Doxorubicin-Induced Cardiotoxicity: Antioxidant Properties and Cell Cycle Modulation. *Int. J. Mol. Sci.* 2023, 24, 12735. [CrossRef]
- 139. Maiuolo, J.; Musolino, V.; Guarnieri, L.; Macrì, R.; Coppoletta, A.R.; Cardamone, A.; Serra, M.; Gliozzi, M.; Bava, I.; Lupia, C.; et al. *Ferula communis* L. (*Apiaceae*) Root Acetone-Water Extract: Phytochemical Analysis, Cytotoxicity and In Vitro Evaluation of Estrogenic Properties. *Plants* 2022, *11*, 1905. [CrossRef] [PubMed]
- 140. Maiuolo, J.; Miceli, N.; Davì, F.; Bava, I.; Tucci, L.; Ragusa, S.; Taviano, M.F.; Musolino, V.; Gliozzi, M.; Carresi, C.; et al. *Ferula communis* Root Extract: In Vitro Evaluation of the Potential Additive Effect with Chemotherapy Tamoxifen in Breast Cancer (MCF-7) Cells Part II. *Plants* 2023, 12, 1194. [CrossRef] [PubMed]
- 141. Safi, R.; Hamade, A.; Bteich, N.; El Saghir, J.; Assaf, M.D.; El-Sabban, M.; Najjar, F. A ferutinin analogue with enhanced potency and selectivity against ER-positive breast cancer cells in vitro. *Biomed. Pharmacother.* **2018**, 105, 267–273. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 37 of 41

142. Maiuolo, J.; Bava, I.; Carresi, C.; Gliozzi, M.; Musolino, V.; Scicchitano, M.; Macri, R.; Oppedisano, F.; Scarano, F.; Caterina Zito, M.; et al. The Effect of *Ferula communis* Extract in *Escherichia coli* Lipopolysaccharide-Induced Neuroinflammation in Cultured Neurons and Oligodendrocytes. *Int. J. Mol. Sci.* 2021, 22, 7910. [CrossRef] [PubMed]

- 143. Adorisio, S.; Muscari, I.; Fierabracci, A.; Thi Thuy, T.; Marchetti, M.C.; Ayroldi, E.; Delfino, D.V. Biological effects of bergamot and its potential therapeutic use as an anti-inflammatory, antioxidant, and anticancer agent. *Pharm. Biol.* **2023**, *61*, 639–646. [CrossRef] [PubMed]
- 144. Tripathi, S.; Sharma, Y.; Kumar, D. Unveiling the link between chronic inflammation and cancer. *Metabol. Open.* **2025**, 25, 100347. [CrossRef] [PubMed]
- 145. Xie, Y.; Liu, F.; Wu, Y.; Zhu, Y.; Jiang, Y.; Wu, Q.; Dong, Z.; Liu, K. Inflammation in cancer: Therapeutic opportunities from new insights. *Mol. Cancer* 2025, 24, 51. [CrossRef] [PubMed]
- 146. Wang, M.; Zhang, L.; Chang, W.; Zhang, Y. The crosstalk between the gut microbiota and tumor immunity: Implications for cancer progression and treatment outcomes. *Front. Immunol.* **2023**, *13*, 1096551. [CrossRef] [PubMed]
- 147. Chen, L.; Deng, H.; Cui, H.; Fang, J.; Zuo, Z.; Deng, J.; Li, Y.; Wang, X.; Zhao, L. Inflammatory responses and inflammation-associated diseases in organs. *Oncotarget* 2017, *9*, 7204–7218. [CrossRef] [PubMed]
- 148. Zhao, H.; Wu, L.; Yan, G.; Chen, Y.; Zhou, M.; Wu, Y.; Li, Y. Inflammation and tumor progression: Signaling pathways and targeted intervention. *Signal Transduct. Target. Ther.* **2021**, *6*, 263. [CrossRef] [PubMed]
- 149. Zhu, A.S.; Mustafa, T.; Connell, J.P.; Grande-Allen, K.J. Tumor necrosis factor alpha and interleukin 1 beta suppress myofibroblast activation via nuclear factor kappa B signaling in 3D-cultured mitral valve interstitial cells. *Acta Biomater.* **2021**, 127, 159–168. [CrossRef] [PubMed]
- 150. Ait-Ahmed, Y.; Lafdil, F. Novel insights into the impact of liver inflammatory responses on primary liver cancer development. *Liver Res.* **2023**, *7*, 26–34. [CrossRef] [PubMed]
- 151. Nishida, A.; Andoh, A. The Role of Inflammation in Cancer: Mechanisms of Tumor Initiation, Progression, and Metastasis. *Cells* **2025**, *14*, 488. [CrossRef] [PubMed]
- 152. Tristan Asensi, M.; Napoletano, A.; Sofi, F.; Dinu, M. Low-Grade Inflammation and Ultra-Processed Foods Consumption: A Review. *Nutrients* 2023, 15, 1546. [CrossRef] [PubMed]
- 153. Minarovits, J. Human tumor viruses: Induction of three-dimensional alterations in the host genome structure. *Front. Microbiol.* **2023**, *14*, 1280210. [CrossRef] [PubMed]
- 154. Sun, M.; Zhu, S.; Wang, Y.; Zhao, Y.; Yan, K.; Li, X.; Wang, X.; He, C.; Ding, C.; Chen, Y.; et al. Effect of inflammation on association between cancer and coronary artery disease. *BMC Cardiovasc. Disord.* 2024, 24, 72. [CrossRef] [PubMed]
- 155. Mora Barthelmess, R.; Stijlemans, B.; Van Ginderachter, J.A. Hallmarks of Cancer Affected by the MIF Cytokine Family. *Cancers* **2023**, *15*, 395. [CrossRef] [PubMed]
- 156. Gao, S.P.; Mark, K.G.; Leslie, K.; Pao, W.; Motoi, N.; Gerald, W.L.; Travis, W.D.; Bornmann, W.; Veach, D.; Clarkson, B.; et al. Mutations in the EGFR kinase domain mediate STAT3 activation via IL-6 production in human lung adenocarcinomas. *J. Clin. Investig.* 2007, 117, 3846–3856. [CrossRef] [PubMed]
- 157. Leone, P.; Malerba, E.; Susca, N.; Favoino, E.; Perosa, F.; Brunori, G.; Prete, M.; Racanelli, V. Endothelial cells in tumor microenvironment: Insights and perspectives. *Front. Immunol.* **2024**, *15*, 1367875. [CrossRef] [PubMed]
- 158. Xiao, Y.; Yu, D. Tumor microenvironment as a therapeutic target in cancer. *Pharmacol. Ther.* **2021**, 221, 107753. [CrossRef] [PubMed]
- 159. Vallée, A.; Lecarpentier, Y. Crosstalk Between Peroxisome Proliferator-Activated Receptor Gamma and the Canonical WNT/β-Catenin Pathway in Chronic Inflammation and Oxidative Stress During Carcinogenesis. *Front. Immunol.* **2018**, *9*, 745. [CrossRef] [PubMed]
- 160. Sabatino, L.; Pancione, M.; Votino, C.; Colangelo, T.; Lupo, A.; Novellino, E.; Lavecchia, A.; Colantuoni, V. Emerging role of the β-catenin-PPARγ axis in the pathogenesis of colorectal cancer. *World J. Gastroenterol.* **2014**, *20*, 7137–7151. [CrossRef] [PubMed]
- 161. Chi, T.; Wang, M.; Wang, X.; Yang, K.; Xie, F.; Liao, Z.; Wei, P. PPAR-γ Modulators as Current and Potential Cancer Treatments. *Front. Oncol.* **2021**, *11*, 737776. [CrossRef] [PubMed]
- 162. Hernandez-Quiles, M.; Broekema, M.F.; Kalkhoven, E. PPARgamma in Metabolism, Immunity, and Cancer: Unified and Diverse Mechanisms of Action. *Front. Endocrinol.* **2021**, 12, 624112. [CrossRef] [PubMed]
- 163. Kurowska, A.; Ziemichód, W.; Herbet, M.; Piątkowska-Chmiel, I. The Role of Diet as a Modulator of the Inflammatory Process in the Neurological Diseases. *Nutrients* **2023**, *15*, 1436. [CrossRef] [PubMed]
- 164. Russo, C.; Lombardo, G.E.; Bruschetta, G.; Rapisarda, A.; Maugeri, A.; Navarra, M. Bergamot Byproducts: A Sustainable Source to Counteract Inflammation. *Nutrients* **2024**, *16*, 259. [CrossRef] [PubMed]
- 165. Carresi, C.; Mollace, R.; Macrì, R.; Scicchitano, M.; Bosco, F.; Scarano, F.; Coppoletta, A.R.; Guarnieri, L.; Ruga, S.; Zito, M.C.; et al. Oxidative Stress Triggers Defective Autophagy in Endothelial Cells: Role in Atherothrombosis Development. *Antioxidants* **2021**, 10, 387. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 38 of 41

166. Parafati, M.; Lascala, A.; La Russa, D.; Mignogna, C.; Trimboli, F.; Morittu, V.M.; Riillo, C.; Macirella, R.; Mollace, V.; Brunelli, E.; et al. Bergamot Polyphenols Boost Therapeutic Effects of the Diet on Non-Alcoholic Steatohepatitis (NASH) Induced by "Junk Food": Evidence for Anti-Inflammatory Activity. *Nutrients* 2018, 10, 1604. [CrossRef] [PubMed]

- 167. Gallyas, F., Jr.; Sumegi, B. Mitochondrial Protection by PARP Inhibition. Int. J. Mol. Sci. 2020, 21, 2767. [CrossRef] [PubMed]
- 168. Sun, J.; Yu, L.; Qu, X.; Huang, T. The role of peroxisome proliferator-activated receptors in the tumor microenvironment, tumor cell metabolism, and anticancer therapy. *Front. Pharmacol.* **2023**, *14*, 1184794. [CrossRef] [PubMed]
- 169. Chen, S.; Liu, Z.; Wu, H.; Wang, B.; Ouyang, Y.; Liu, J.; Zheng, X.; Zhang, H.; Li, X.; Feng, X.; et al. Adipocyte-rich microenvironment promotes chemoresistance via upregulation of peroxisome proliferator-activated receptor gamma/ABCG2 in epithelial ovarian cancer. *Int. J. Mol. Med.* 2024, 53, 37. [CrossRef] [PubMed]
- 170. Nakamura, T.; Pitna, D.B.; Kimura, K.; Yoshimoto, Y.; Uchiyama, T.; Mori, T.; Kondo, R.; Hara, S.; Egoshi, Y.; Yamaguchi, S.; et al. Total synthesis of cynaropicrin. *Org. Biomol. Chem.* **2021**, *19*, 6038–6044. [CrossRef] [PubMed]
- 171. Chen, J.; Liu, X. Cynaropicrin attenuates inflammatory cytokines in LPS-induced RAW264.7 cells and ovalbumin-induced asthmatic mice. *J. Biochem. Mol. Toxicol.* **2024**, *38*, e23836. [CrossRef] [PubMed]
- 172. Silvestrini, A.; Giordani, C.; Bonacci, S.; Giuliani, A.; Ramini, D.; Matacchione, G.; Sabbatinelli, J.; Di Valerio, S.; Pacetti, D.; Procopio, A.D.; et al. Anti-Inflammatory Effects of Olive Leaf Extract and Its Bioactive Compounds Oleacin and Oleuropein-Aglycone on Senescent Endothelial and Small Airway Epithelial Cells. *Antioxidants* 2023, 12, 1509. [CrossRef] [PubMed]
- 173. Tamburini, B.; Di Liberto, D.; Pratelli, G.; Rizzo, C.; Barbera, L.; Lauricella, M.; Carlisi, D.; Maggio, A.; Palumbo Piccionello, A.; D'Anneo, A.; et al. Extra Virgin Olive Oil Polyphenol-Enriched Extracts Exert Antioxidant and Anti-Inflammatory Effects on Peripheral Blood Mononuclear Cells from Rheumatoid Arthritis Patients. *Antioxidants* 2025, 14, 171. [CrossRef] [PubMed]
- 174. Mirsanei, Z.; Heidari, N.; Hazrati, A.; Asemani, Y.; Niknam, B.; Yousefi, Z.; Jafari, R. Oleuropein reduces LPS-induced inflammation via stimulating M2 macrophage polarization. *Biomed. Pharmacother.* **2023**, *163*, 114857. [CrossRef] [PubMed]
- 175. Ruzzolini, J.; Chioccioli, S.; Monaco, N.; Peppicelli, S.; Andreucci, E.; Urciuoli, S.; Romani, A.; Luceri, C.; Tortora, K.; Calorini, L.; et al. Oleuropein-Rich Leaf Extract as a Broad Inhibitor of Tumour and Macrophage iNOS in an Apc Mutant Rat Model. *Antioxidants* 2021, 10, 1577. [CrossRef] [PubMed]
- 176. Bigagli, E.; Cinci, L.; Paccosi, S.; Parenti, A.; D'Ambrosio, M.; Luceri, C. Nutritionally relevant concentrations of resveratrol and hydroxytyrosol mitigate oxidative burst of human granulocytes and monocytes and the production of pro-inflammatory mediators in LPS-stimulated RAW 264.7 macrophages. *Int. Immunopharmacol.* 2017, 43, 147–155. [CrossRef] [PubMed]
- 177. Jin, H.X.; Zhang, Y.H.; Guo, R.N.; Zhao, S.N. Inhibition of MEK/ERK/STAT3 signaling in oleuropein treatment inhibits myocardial ischemia/reperfusion. *Int. J. Mol. Med.* **2018**, 42, 1034–1043. [CrossRef] [PubMed]
- 178. Karakoç, M.D.; Sekkin, S. Effects of Oleuropein on Epirubicin and Cyclophosphamide Combination Treatment in Rats. *Turk. J. Pharm. Sci.* **2021**, *18*, 420–429. [CrossRef] [PubMed]
- 179. Vezza, T.; Rodríguez-Nogales, A.; Algieri, F.; Garrido-Mesa, J.; Romero, M.; Sánchez, M.; Toral, M.; Martín-García, B.; Gómez-Caravaca, A.M.; Arráez-Román, D.; et al. The metabolic and vascular protective effects of olive (*Olea europaea* L.) leaf extract in diet-induced obesity in mice are related to the amelioration of gut microbiota dysbiosis and to its immunomodulatory properties. *Pharmacol. Res.* **2019**, *150*, 104487. [CrossRef] [PubMed]
- 180. Rehman, A.; Bukhari, S.A.; Chauhdary, Z.; Akhter, N.; Noreen, R. Effect of resveratrol and quercetin on SFRP4 as a biomarker of diabesity: In silico and in vivo studies. *Biochem. Biophys. Res. Commun.* **2025**, 761, 151748. [CrossRef] [PubMed]
- 181. Zhao, L.; Cen, F.; Tian, F.; Li, M.J.; Zhang, Q.; Shen, H.Y.; Shen, X.C.; Zhou, M.M.; Du, J. Combination treatment with quercetin and resveratrol attenuates high fat diet-induced obesity and associated inflammation in rats via the AMPKα1/SIRT1 signaling pathway. *Exp. Ther. Med.* **2017**, *14*, 5942–5948. [CrossRef] [PubMed]
- 182. Li, L.; Jiang, W.; Yu, B.; Liang, H.; Mao, S.; Hu, X.; Feng, Y.; Xu, J.; Chu, L. Quercetin improves cerebral ischemia/reperfusion injury by promoting microglia/macrophages M2 polarization via regulating PI3K/Akt/NF-κB signaling pathway. *Biomed. Pharmacother.* **2023**, *168*, 115653. [CrossRef] [PubMed]
- 183. Ma, E.; Wu, C.; Chen, J.; Wo, D.; Ren, D.N.; Yan, H.; Peng, L.; Zhu, W. Resveratrol prevents Ang II-induced cardiac hypertrophy by inhibition of NF-κB signaling. *Biomed. Pharmacother.* **2023**, *165*, 115275. [CrossRef] [PubMed]
- 184. Geroushi, A.; Auzi, A.A.; Elhwuegi, A.S.; Elzawam, F.; Elsherif, A.; Nahar, L.; Sarker, S.D. Antiinflammatory sesquiterpenes from the root oil of Ferula hermonis. *Phytother. Res.* **2011**, 25, 774–777. [CrossRef] [PubMed]
- 185. Zhang, M.; Zhang, L.; Hei, R.; Li, X.; Cai, H.; Wu, X.; Zheng, Q.; Cai, C. CDK inhibitors in cancer therapy, an overview of recent development. *Am. J. Cancer Res.* **2021**, *11*, 1913–1935. [PubMed]
- 186. Ozaki, T.; Nakagawara, A. Role of p53 in Cell Death and Human Cancers. Cancers 2011, 3, 994–1013. [CrossRef] [PubMed]
- 187. Fan, W.; Xiong, R.; Zhou, Z.; Zhang, C.; Han, Y.; Shi, T.; Qiu, J.; Zhang, R. ZFP57 promotes ovarian cancer progression by transcriptionally regulating BRCA1 and managing G1 checkpoint. *J. Cancer* 2023, 14, 2039–2050. [CrossRef] [PubMed]
- 188. Liu, Y.; Liang, W.; Chang, Y.; He, Z.; Wu, M.; Zheng, H.; Ke, X.; Lv, M.; Liu, Q.; Liu, Q.; et al. CEP192 is a novel prognostic marker and correlates with the immune microenvironment in hepatocellular carcinoma. *Front. Immunol.* **2022**, *13*, 950884. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 39 of 41

189. Gheghiani, L.; Wang, L.; Zhang, Y.; Moore, X.T.R.; Zhang, J.; Smith, S.C.; Tian, Y.; Wang, L.; Turner, K.; Jackson-Cook, C.K.; et al. PLK1 Induces Chromosomal Instability and Overrides Cell-Cycle Checkpoints to Drive Tumorigenesis. *Cancer Res.* **2021**, *81*, 1293–1307. [CrossRef] [PubMed]

- 190. Corno, A.; Cordeiro, M.H.; Allan, L.A.; Lim, Q.W.; Harrington, E.; Smith, R.J.; Saurin, A.T. A bifunctional kinase-phosphatase module balances mitotic checkpoint strength and kinetochore-microtubule attachment stability. *EMBO J.* **2023**, *42*, 112630. [CrossRef] [PubMed]
- 191. Łukasik, P.; Załuski, M.; Gutowska, I. Cyclin-Dependent Kinases (CDK) and Their Role in Diseases Development-Review. *Int. J. Mol. Sci.* **2021**, 22, 2935. [CrossRef] [PubMed]
- 192. Boccardi, V.; Marano, L. Aging, Cancer, and Inflammation: The Telomerase Connection. *Int. J. Mol. Sci.* **2024**, *25*, 8542. [CrossRef] [PubMed]
- 193. Dietrich, C.; Trub, A.; Ahn, A.; Taylor, M.; Ambani, K.; Chan, K.T.; Lu, K.H.; Mahendra, C.A.; Blyth, C.; Coulson, R.; et al. INX-315, a Selective CDK2 Inhibitor, Induces Cell Cycle Arrest and Senescence in Solid Tumors. *Cancer Discov.* **2024**, *14*, 446–467. [CrossRef] [PubMed]
- 194. Singh, S.R.; Bhaskar, R.; Ghosh, S.; Yarlagadda, B.; Singh, K.K.; Verma, P.; Sengupta, S.; Mladenov, M.; Hadzi-Petrushev, N.; Stojchevski, R.; et al. Exploring the Genetic Orchestra of Cancer: The Interplay Between Oncogenes and Tumor-Suppressor Genes. *Cancers* 2025, 17, 1082. [CrossRef] [PubMed]
- 195. Garcia-Oliveira, P.; Otero, P.; Pereira, A.G.; Chamorro, F.; Carpena, M.; Echave, J.; Fraga-Corral, M.; Simal-Gandara, J.; Prieto, M.A. Status and Challenges of Plant-Anticancer Compounds in Cancer Treatment. *Pharmaceuticals* **2021**, *14*, 157. [CrossRef] [PubMed]
- 196. Maugeri, A.; Russo, C.; Musumeci, L.; Lombardo, G.E.; De Sarro, G.; Barreca, D.; Cirmi, S.; Navarra, M. The Anticancer Effect of a Flavonoid-Rich Extract of Bergamot Juice in THP-1 Cells Engages the SIRT2/AKT/p53 Pathway. *Pharmaceutics* **2022**, *14*, 2168. [CrossRef]
- 197. Delle Monache, S.; Sanità, P.; Trapasso, E.; Ursino, M.R.; Dugo, P.; Russo, M.; Ferlazzo, N.; Calapai, G.; Angelucci, A.; Navarra, M. Mechanisms underlying the anti-tumoral effects of Citrus Bergamia juice. *PLoS ONE* **2013**, *8*, 61484. [CrossRef] [PubMed]
- 198. Navarra, M.; Femia, A.P.; Romagnoli, A.; Tortora, K.; Luceri, C.; Cirmi, S.; Ferlazzo, N.; Caderni, G. A flavonoid-rich extract from bergamot juice prevents carcinogenesis in a genetic model of colorectal cancer, the Pirc rat (F344/NTac-Apc^{am1137}). *Eur. J. Nutr.* **2020**, *59*, 885–894. [CrossRef] [PubMed]
- 199. Lepore, S.M.; Maggisano, V.; Lombardo, G.E.; Maiuolo, J.; Mollace, V.; Bulotta, S.; Russo, D.; Celano, M. Antiproliferative Effects of Cynaropicrin on Anaplastic Thyroid Cancer Cells. *Endocr. Metab. Immune Disord. Drug Targets* **2019**, *19*, 59–66. [CrossRef] [PubMed]
- 200. Ding, Z.; Xi, J.; Zhong, M.; Chen, F.; Zhao, H.; Zhang, B.; Fang, J. Cynaropicrin Induces Cell Cycle Arrest and Apoptosis by Inhibiting PKM2 to Cause DNA Damage and Mitochondrial Fission in A549 Cells. J. Agric. Food Chem. 2021, 69, 13557–13567. [CrossRef] [PubMed]
- 201. Pessoa, H.R.; Zago, L.; Difonzo, G.; Pasqualone, A.; Caponio, F.; Ferraz da Costa, D.C. Olive Leaves as a Source of Anticancer Compounds: In Vitro Evidence and Mechanisms. *Molecules* **2024**, *29*, 4249. [CrossRef] [PubMed]
- 202. Han, S.; Hou, H.; Zhang, Y.; Fang, Z.; Li, Y.; Zhang, L.; Wang, Y.; Zhang, S.; Zhang, H.; Jin, Q.; et al. Oleuropein and its hydrolysate from extra virgin olive oil inhibit breast cancer cells proliferation interfering with the PI3K-AKT signal pathway. *J. Funct. Foods* 2023, 110, 1756–4646. [CrossRef]
- 203. Messeha, S.S.; Zarmouh, N.O.; Asiri, A.; Soliman, K.F.A. Gene Expression Alterations Associated with Oleuropein-Induced Antiproliferative Effects and S-Phase Cell Cycle Arrest in Triple-Negative Breast Cancer Cells. *Nutrients* 2020, 12, 3755. [CrossRef] [PubMed]
- 204. Benitez, D.A.; Pozo-Guisado, E.; Alvarez-Barrientos, A.; Fernandez-Salguero, P.M.; Castellón, E.A. Mechanisms involved in resveratrol-induced apoptosis and cell cycle arrest in prostate cancer-derived cell lines. *J. Androl.* 2007, 28, 282–293. [CrossRef] [PubMed]
- 205. Ahmad, N.; Adhami, V.M.; Afaq, F.; Feyes, D.K.; Mukhtar, H. Resveratrol causes WAF-1/p21-mediated G(1)-phase arrest of cell cycle and induction of apoptosis in human epidermoid carcinoma A431 cells. *Clin. Cancer Res.* **2001**, *7*, 1466–1473. [PubMed]
- 206. Bailon-Moscoso, N.; Cevallos-Solorzano, G.; Romero-Benavides, J.C.; Orellana, M.I. Natural Compounds as Modulators of Cell Cycle Arrest: Application for Anticancer Chemotherapies. *Curr. Genom.* **2017**, *18*, 106–131. [CrossRef] [PubMed]
- 207. Wu, H.; Chen, L.; Zhu, F.; Han, X.; Sun, L.; Chen, K. The Cytotoxicity Effect of Resveratrol: Cell Cycle Arrest and Induced Apoptosis of Breast Cancer 4T1 Cells. *Toxins* 2019, 11, 731. [CrossRef] [PubMed]
- 208. Shi, Y.; Cai, G.; Zhang, C.; Li, H.; Nie, Y.; Yu, S.; Zhang, B.; Wu, M.; Luo, W.; Liu, J.; et al. Resveratrol suppresses growth and VCAN expression in a Cancer-associated fibroblast-breast Cancer hybrid organoid. *Int. Immunopharmacol.* **2025**, *153*, 114451. [CrossRef] [PubMed]
- 209. Catanzaro, D.; Ragazzi, E.; Vianello, C.; Caparrotta, L.; Montopoli, M. Effect of Quercetin on Cell Cycle and Cyclin Expression in Ovarian Carcinoma and Osteosarcoma Cell Lines. *Nat. Prod. Commun.* 2015, 10, 1365–1368. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 40 of 41

210. Suh, D.K.; Lee, E.J.; Kim, H.C.; Kim, J.H. Induction of G(1)/S phase arrest and apoptosis by quercetin in human osteosarcoma cells. *Arch. Pharm. Res.* **2010**, *33*, 781–785. [CrossRef] [PubMed]

- 211. Son, H.K.; Kim, D. Quercetin Induces Cell Cycle Arrest and Apoptosis in YD10B and YD38 Oral Squamous Cell Carcinoma Cells. *Asian Pac. J. Cancer Prev.* 2023, 24, 283–289. [CrossRef] [PubMed]
- 212. Hussein, S.A.; Ababneh, N.A.; Tarawneh, N.; Ismail, M.A.; Awidi, A.; Abdalla, S. Antitumor Effects of Quercetin and Luteolin in A375 Cutaneous Melanoma Cell Line Are Mediated by Upregulation of P-ERK, c-Myc, and the Upstream GPER. *Life* 2025, 15, 417. [CrossRef] [PubMed]
- 213. Keshmirshekan, F.; Mohamadi-Zarch, S.M.; Bagheri, S.M. A Review of Anticancer Potential of Conferone, Diversin and Ferutinin; Which One is Stronger for Cancer Therapy? *Anti-Cancer Agents Med. Chem.* **2025**, 25, 378–387. [CrossRef] [PubMed]
- 214. Sasso, J.M.; Ammar, R.M.; Tenchov, R.; Lemmel, S.; Kelber, O.; Grieswelle, M.; Zhou, Q.A. Gut Microbiome-Brain Alliance: A Landscape View into Mental and Gastrointestinal Health and Disorders. *ACS Chem. Neurosci.* 2023, 14, 1717–1763. [CrossRef] [PubMed]
- 215. Zhai, B.; Ola, M.; Rolling, T.; Tosini, N.L.; Joshowitz, S.; Littmann, E.R.; Amoretti, L.A.; Fontana, E.; Wright, R.J.; Miranda, E.; et al. High-resolution mycobiota analysis reveals dynamic intestinal translocation preceding invasive candidiasis. *Nat. Med.* **2020**, *26*, 59–64. [CrossRef] [PubMed]
- 216. Dohlman, A.B.; Klug, J.; Mesko, M.; Gao, I.H.; Lipkin, S.M.; Shen, X.; Iliev, I.D. A pan-cancer mycobiome analysis reveals fungal involvement in gastrointestinal and lung tumors. *Cell* **2022**, *185*, 3807–3822.e12. [CrossRef] [PubMed]
- 217. Galeano Niño, J.L.; Wu, H.; LaCourse, K.D.; Kempchinsky, A.G.; Baryiames, A.; Barber, B.; Futran, N.; Houlton, J.; Sather, C.; Sicinska, E.; et al. Effect of the intratumoral microbiota on spatial and cellular heterogeneity in cancer. *Nature* **2022**, *611*, 810–817. [CrossRef] [PubMed]
- 218. Cao, Y.; Xia, H.; Tan, X.; Shi, C.; Ma, Y.; Meng, D.; Zhou, M.; Lv, Z.; Wang, S.; Jin, Y. Intratumoural microbiota: A new frontier in cancer development and therapy. *Signal Transduct. Target. Ther.* **2024**, *9*, 15. [CrossRef] [PubMed]
- 219. Liu, G.; Liu, K.; Ji, L.; Li, Y. Intratumoral microbiota, fatty acid metabolism, and tumor microenvironment constitute an unresolved trinity in colon adenocarcinoma. *Sci. Rep.* **2025**, *15*, 2568. [CrossRef] [PubMed]
- 220. Ruiz-Malagón, A.J.; Rodríguez-Sojo, M.J.; Redondo, E.; Rodríguez-Cabezas, M.E.; Gálvez, J.; Rodríguez-Nogales, A. Systematic review: The gut microbiota as a link between colorectal cancer and obesity. *Obes. Rev.* 2025, 26, 13872. [CrossRef] [PubMed]
- 221. Xu, J.; Cheng, M.; Liu, J.; Cui, M.; Yin, B.; Liang, J. Research progress on the impact of intratumoral microbiota on the immune microenvironment of malignant tumors and its role in immunotherapy. *Front. Immunol.* **2024**, *15*, 1389446. [CrossRef] [PubMed]
- 222. Hu, Z.J.; Zhu, H.R.; Jin, Y.J.; Liu, P.; Yu, X.W.; Zhang, Y.R. Correlation between gut microbiota and tumor immune microenvironment: A bibliometric and visualized study. *World J. Clin. Oncol.* **2025**, *16*, 101611. [CrossRef] [PubMed]
- 223. Jovel, J.; Patterson, J.; Wang, W.; Hotte, N.; O'Keefe, S.; Mitchel, T.; Perry, T.; Kao, D.; Mason, A.L.; Madsen, K.L.; et al. Characterization of the Gut Microbiome Using 16S or Shotgun Metagenomics. *Front. Microbiol.* 2016, 7, 459. [CrossRef] [PubMed]
- 224. Wensel, C.R.; Pluznick, J.L.; Salzberg, S.L.; Sears, C.L. Next-generation sequencing: Insights to advance clinical investigations of the microbiome. *J. Clin. Investig.* 2022, 132, 154944. [CrossRef] [PubMed]
- 225. Zhang, Y.J.; Li, S.; Gan, R.Y.; Zhou, T.; Xu, D.P.; Li, H.B. Impacts of gut bacteria on human health and diseases. *Int. J. Mol. Sci.* 2015, 16, 7493–7519. [CrossRef] [PubMed]
- 226. Weersma, R.K.; Zhernakova, A.; Fu, J. Interaction between drugs and the gut microbiome. *Gut* **2020**, *69*, 1510–1519. [CrossRef] [PubMed]
- 227. Riehl, L.; Fürst, J.; Kress, M.; Rykalo, N. The importance of the gut microbiome and its signals for a healthy nervous system and the multifaceted mechanisms of neuropsychiatric disorders. *Front. Neurosci.* **2024**, *17*, 1302957. [CrossRef] [PubMed]
- 228. Sun, J.; Chen, F.; Wu, G. Potential effects of gut microbiota on host cancers: Focus on immunity, DNA damage, cellular pathways, and anticancer therapy. *ISME J.* 2023, 17, 1535–1551. [CrossRef] [PubMed]
- 229. He, R.; Qi, P.; Shu, L.; Ding, Y.; Zeng, P.; Wen, G.; Xiong, Y.; Deng, H. Dysbiosis and extraintestinal cancers. *J. Exp. Clin. Cancer Res.* **2025**, *44*, 44. [CrossRef] [PubMed]
- 230. Zhang, P. Influence of Foods and Nutrition on the Gut Microbiome and Implications for Intestinal Health. *Int. J. Mol. Sci.* **2022**, 23, 9588. [CrossRef] [PubMed]
- 231. Tang, Y.; Cai, Q.; Tian, Z.; Chen, W.; Tang, H. Crosstalk between Gut Microbiota and Cancer Immunotherapy: Present Investigations and Future Perspective. *Research* 2025, 8, 0600. [CrossRef] [PubMed]
- 232. McGill, C.R.; Fulgoni, V.L., 3rd; Devareddy, L. Ten-year trends in fiber and whole grain intakes and food sources for the United States population: National Health and Nutrition Examination Survey 2001–2010. *Nutrients* 2015, 7, 1119–1130. [CrossRef] [PubMed]
- 233. Schnorr, S.L.; Candela, M.; Rampelli, S.; Centanni, M.; Consolandi, C.; Basaglia, G.; Turroni, S.; Biagi, E.; Peano, C.; Severgnini, M.; et al. Gut microbiome of the Hadza hunter-gatherers. *Nat. Commun.* **2014**, *5*, 3654. [CrossRef] [PubMed]

Nutrients **2025**, 17, 2354 41 of 41

234. Wang, D.D.; Qi, Q.; Wang, Z.; Usyk, M.; Sotres-Alvarez, D.; Mattei, J.; Tamez, M.; Gellman, M.D.; Daviglus, M.; Hu, F.B.; et al. The Gut Microbiome Modifies the Association Between a Mediterranean Diet and Diabetes in USA Hispanic/Latino Population. *J. Clin. Endocrinol. Metab.* 2022, 107, e924–e934. [CrossRef] [PubMed]

- 235. Zeb, F.; Naqeeb, H.; Osaili, T.; Faris, M.E.; Ismail, L.C.; Obaid, R.S.; Naja, F.; Radwan, H.; Hasan, H.; Hashim, M.; et al. Molecular crosstalk between polyphenols and gut microbiota in cancer prevention. *Nutr. Res.* **2024**, *124*, 21–42. [CrossRef] [PubMed]
- 236. Li, C.; Xu, Y.; Zhang, J.; Zhang, Y.; He, W.; Ju, J.; Wu, Y.; Wang, Y. The effect of resveratrol, curcumin and quercetin combination on immuno-suppression of tumor microenvironment for breast tumor-bearing mice. *Sci. Rep.* 2023, *13*, 13278. [CrossRef] [PubMed]
- 237. Mandle, H.B.; Jenab, M.; Gunter, M.J.; Tjønneland, A.; Olsen, A.; Dahm, C.C.; Zhang, J.; Sugier, P.E.; Rothwell, J.; Severi, G.; et al. Inflammation and gut barrier function-related genes and colorectal cancer risk in western European populations. *Mutagenesis* 2025, 40, 48–60. [CrossRef] [PubMed]
- 238. Ke, J.; Zhang, C.J.; Wang, L.Z.; Xie, F.S.; Wu, H.Y.; Li, T.; Bian, C.W.; Wu, R.L. Lipopolysaccharide promotes cancer cell migration and invasion through METTL3/PI3K/AKT signaling in human cholangiocarcinoma. *Heliyon* **2024**, *10*, 29683. [CrossRef] [PubMed]
- 239. Mollace, A.; Coluccio, M.L.; Donato, G.; Mollace, V.; Malara, N. Cross-talks in colon cancer between RAGE/AGEs axis and inflammation/immunotherapy. *Oncotarget* **2021**, *12*, 1281–1295. [CrossRef] [PubMed]
- 240. Ramamoorthy Rajalakshmi, R.; Melians, M.A.; Pon, F.F.; Cosio, D.S.; Buvarahamurthy, V.; Jayakumar, A.R.; Paidas, M.J. *Advantages and Disadvantages of Nutraceuticals*; Springer Nature: Singapore, 2023; pp. 245–286. [CrossRef]
- 241. Shaman, J.A. The Future of Pharmacogenomics: Integrating Epigenetics, Nutrigenomics, and Beyond. *J. Pers. Med.* **2024**, *14*, 1121. [CrossRef] [PubMed]
- 242. Zhang, B.; Zhang, Y.; Xing, X.; Wang, S. Health benefits of dietary polyphenols: Insight into interindividual variability in absorption and metabolism. *Curr. Opin. Food Sci.* **2022**, *48*, 2214–7993. [CrossRef]
- 243. Favari, C.; Rinaldi de Alvarenga, J.F.; Sánchez-Martínez, L.; Tosi, N.; Mignogna, C.; Cremonini, E.; Manach, C.; Bresciani, L.; Del Rio, D.; Mena, P. Factors driving the inter-individual variability in the metabolism and bioavailability of (poly)phenolic metabolites: A systematic review of human studies. *Redox Biol.* 2024, 71, 103095. [CrossRef] [PubMed]
- 244. Sahraeian, S.; Rashidinejad, A.; Golmakani, M.-T. Recent advances in the conjugation approaches for enhancing the bioavailability of polyphenols. *Food Hydrocoll.* **2024**, *146*, 109221. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.